

1 CUTTING LENGTH AND BUD REMOVAL ON CASSAVA YIELD

2
3 **EFFECTS OF CUTTING LENGTH AND BUD REMOVAL ON ROOT YIELD AND**
4 **STARCH CONTENT OF CASSAVA UNDER RAINFED CONDITIONS**

5
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14
15 SUMMARY

16
17 Bud removal of the cuttings at underground level has been claimed by cassava growers in Thailand
18 as a method to increase cassava yield. This practice should be tested experimentally to explain the
19 reason for yield increase. The objective of this study was to investigate the effects of bud removal
20 and cutting length on storage root yield and starch content of three cassava varieties. Field
21 experiment was conducted in a split-split plot design with four replications in 2010 and 2011,
22 under rainfed conditions. Three cassava varieties (KU50, RY9 and HB60) were assigned as main
23 plot. Two cutting lengths (15 cm and 30 cm) were assigned as sub plots, and two treatments of
24 buds (buds cut and not cut) were assigned as sub-sub plots. The buds on the cuttings that were
25 inserted into the soil were removed. In 2010, the plants from 15 cm long cuttings subjected to bud
26 removal had higher fresh storage root yield (88.4 Mg ha⁻¹) than did plants from 30 cm long cuttings
27 subjected to bud removal (75.8 Mg ha⁻¹). Cutting of buds also had higher fresh storage root yield
28 (89.1 Mg ha⁻¹) than did non bud-cutting (75.0 Mg ha⁻¹). KU50 had the highest fresh storage root
29 yield (91.4 Mg ha⁻¹), dry root yield (48.4 Mg ha⁻¹) and starch yield (20.1 Mg ha⁻¹). Cutting length
30 of 15 cm had higher starch concentration in storage roots (25.6%) than did cutting length of 30 cm
31 (24.2%). HB60 had the highest starch concentration (27.0%) among cassava varieties tested. The
32 data in 2011 were similar to the data in 2010. The responses of varieties to bud removal and cutting
33 length are discussed.

34
35 Keywords: bud removal, cutting length, farmers' practice, root yield

INTRODUCTION

Cassava (*Manihot esculenta* Crantz) originated in the tropical areas of South America and it is grown widely in tropical areas of the world due to its starch containing roots (Alves, 2001; Scott *et al.*, 2000). Globally, the area planted with cassava was recently estimated at 20.4 million hectares with a total production of 262.5 million tonnes (MT) (FAO, 2014). Thailand ranks third for world cassava production with an average total production of 22.5 MT year⁻¹, with the areas in the northeastern region accounting for approximately 51.3% of the country's entire cassava production (OAE, 2014). The national average yield in 2006 was about 20 MT ha⁻¹, which is higher than the world average but lower than the potential of the crop (FAO, 2014). The main causal factors driving low yields in Thailand are infertile soils and drought (FAO, 2014), as well as inappropriate agronomic practices (Jones, 2013).

It has been reported previously that cassava yield depends on plant population density, number of roots and tuber root weight per plant (Hahn and Hozyo, 1984). In cassava cultivation, the cuttings from the stems are used as planting materials for the succeeding crop (Nassar and Teixeira, 1983; Leihner, 2002). The number of cassava roots can be increased by adopting a vertical planting position and using longer cuttings (Osiru *et al.*, 1997). However, no significant difference in root yield between cuttings with 10 cm and 50 cm length has been reported (Velasco, 1982). Short cuttings produced better yields than long cuttings (Villamayor *et al.*, 1992) and cuttings with 20 cm length have higher root yield than longer cuttings with 25 cm length (Tongglum *et al.*, 1992).

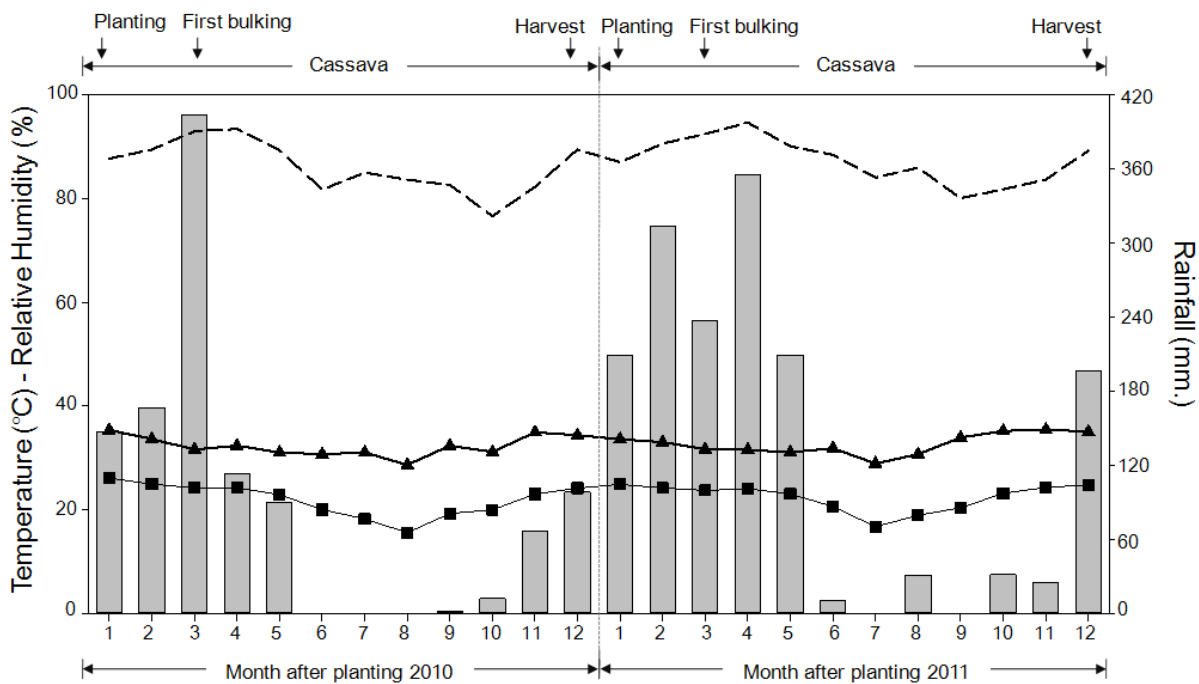
One local agronomic practice that the farmers in northeastern Thailand claim to considerably increase root yield of cassava is to cut the buds off the section of the cuttings to be inserted into the soil. However, the yield components that contribute to root yield have not been clearly investigated. The hypothesis underlying this research is that bud removal increases storage root yield and modifies yield components of cassava. The reason for yield increase of this local practice has not been verified experimentally. Therefore, the objective of this research was to evaluate the effects of cutting length and bud removal on plant growth, root yield and starch content of three cassava varieties grown under rain-fed conditions in northeastern Thailand.

MATERIALS AND METHODS

Crop management and experimental design

Field experiments were conducted at the Agronomy Experimental Fields of Khon Kaen University, Khon Kaen, Thailand (16°28'N, 102°48'E, 200 m above sea level) under rainfed conditions in two consecutive years (2010 and 2011). The crops were planted in June and harvested after 12 months for both years. Soil type is loamy sand in texture. Soil samples were collected from 0-30 cm depth before planting and analyzed for selected chemical and physical properties (Supplementary Material Table S1).

Rainfall, maximum and minimum air temperature were recorded daily and summarized on a monthly basis for entire growing period (Figure 1). Total rainfall in 2010 and 2011 during the growing period was recorded as 1,099 mm and 1,620 mm, respectively. The highest monthly rainfalls observed were 404 mm in August 2010 and 356 mm in September 2011.



81
 82 Figure 1. Rainfall (—), relative humidity (---), minimum temperature (—■—) and maximum
 83 temperature (—▲—) entire the growing period in 2010 and 2011. Khon Kaen, Thailand.

84
 85 The experiment was subjected to water stress during the growing period for 4 months in
 86 2010 (starting at 6 months after planting, MAP) and 3 months in 2011 (Figure 1). The experiment
 87 also encountered intermittent water-logging events for 5 months in 2011, starting at 1 MAP.

88 The experimental fields were ploughed twice with a 3-disk tractor and a 7-disk tractor.
 89 Ridging was undertaken with a 7-disk tractor forming a ridge height of 30 cm. The distance
 90 between planting rows and cassava plants was 1 m. Pre-emergence herbicide (metholachor) was
 91 sprayed immediately after planting at the rate of 1.5 kg a.i. ha⁻¹. A split-split plot design with four
 92 replications and plot size of 11 × 12 m was adopted in this study.

93 Three cassava varieties (Kasetsart 50 [KU50], Rayong 9 [RY9] and Huay Bong 60 [HB60]),
 94 two cutting lengths of 15 cm and 30 cm and two bud removal treatments (bud cut and not cut)
 95 were assigned in main plots, sub-plots and sub-sub-plots, respectively. Three buds on the cutting
 96 of 15 cm length (from six normal buds) and five buds on the cutting of 30 cm length (from ten
 97 normal buds) were removed using a sharp knife. The buds, bark and cambium layer were removed
 98 (Supplementary Material Figure S1). The cuttings were inserted vertically into the soil with two-
 99 third of the length exposed on top of ridges. Chemical fertilizer of grade 15-15-15 (N, P₂O₅, K₂O)
 100 at the rate of 312.5 kg ha⁻¹ (31.3 g plant⁻¹) was applied 1 month after planting. The granule fertilizer
 101 was dropped into the hole made using a hand hoe 15 cm from the cassava plant and covered with
 102 soil.

103 Hand weeding was undertaken only one time at 1 month after planting and no weeding was
104 done for the rest of cropping period in both years. No insecticide or fungicide was used in these
105 experiments for the entire growing period over both years.

106 107 *Data collection*

108 Four plants from each plot were selected randomly outside the harvesting area of the
109 experimental plots at 120, 240, 300 and 360 days after planting (DAP) and shoot dry weight per
110 plant, storage root dry weight per plant and storage root number per plant were evaluated. The
111 harvested plants were separated into leaves, stems and roots. Twenty leaves of each plant from
112 each plot were randomly chosen and leaf area was determined by an automatic leaf area meter
113 (model AAC-400, Hayashi Denkoh Co., Ltd., Bunkyo-ku, Tokyo, Japan). The leaf samples were
114 subsequently oven-dried at 80 °C to a constant weight and leaf dry weight was measured. Leaf dry
115 weight and leaf area of 20 leaves were converted to leaf dry weight and leaf area of total sample
116 using the relationship; total leaf area equals leaf area of 20 leaves × total leaf dry weight/leaf dry
117 weight of 20 leaves. Leaf area index (LAI) was calculated by leaf area per plant divided by ground
118 area covered (Ekanayake, 1994).

119 Fresh storage root yield and dry storage root yield were measured at 360 DAP in the
120 harvesting area of 6 × 4 m (twenty-four plants). Subsequently, fresh and dry storage root yield
121 were calculated in Mg ha⁻¹. Harvest index (HI) was calculated by storage root dry weight divided
122 by total plant dry weight (Ekanayake, 1994). Starch content was measured using a Riemann scale
123 balance (Bainbridge *et al.*, 1996) and starch yield was calculated by multiplying the actual starch
124 content with the fresh root weight per hectare and divided by 100 (Knutsson, 2012).

125 126 *Data analysis*

127 The data for each year were analyzed statistically according to a split-split plot design using
128 Statistica Ver. 11 (Statsoft Inc., Tulsa, USA) and the error variances were tested for variance
129 homogeneity. As some error variances were three-fold different, means were separated by post-
130 hoc Fisher's least significant difference (LSD) at 0.05 probability level.

131 132 RESULTS

133 *Fresh storage root yield, dry storage root yield and harvest index*

134 Cutting lengths affected ($P \leq 0.05$) fresh storage root yield, dry storage root yield and harvest
135 index across years (Table 1). Cutting of 15 cm length caused higher values than that of 30 cm for
136 these traits. Significant differences ($P \leq 0.05$) between bud removal treatments were also observed
137 and cutting with bud removal induced the highest fresh storage root yield, dry storage root yield
138 and harvest index across years (Table 1). Cassava varieties were also significantly different
139 ($P \leq 0.05$) for these traits across years, with KU50 presenting the highest fresh storage root yield
140 (91.4 Mg ha⁻¹ in 2010 and 87.4 Mg ha⁻¹ in 2011), dry storage root yield (48.4 Mg ha⁻¹ in 2010 and
141 44.7 Mg ha⁻¹ in 2011) and harvest index (0.53 in 2010 and 0.52 in 2011).

142 143 *Starch yield and starch content*

144 Significant differences ($P \leq 0.05$) between cutting lengths were observed for starch yield and
145 starch concentration (Table 1). Cutting of 15 cm length determined the highest starch yields (22.6
146 Mg ha⁻¹ in 2010 and 18.5 Mg ha⁻¹ in 2011) and starch concentrations (25.6% in 2010 and 25.1%
147 in 2011). Cutting with bud removal induced the highest ($P \leq 0.05$) starch yields (22.5 Mg ha⁻¹ in
148 2010 and 18.6 Mg ha⁻¹ in 2011) and starch concentration (25.3% in 2010 and 24.8% in 2011).

149 Cassava varieties were also significantly different ($P \leq 0.05$) for starch yield and starch
 150 concentration across years. KU50 had the highest starch yields of 20.1 and 18.7 Mg ha⁻¹ in 2010
 151 and 2011, respectively, whereas HB60 and RY9 had the highest starch concentration of 27.0% and
 152 25.7% in 2010 and 26.5% and 25.2% in 2011, respectively.

153

154 Table 1. Fresh storage root yield (FRY), dry storage root yield (DRY), starch yield (SY), starch
 155 content (SC) and harvest index (HI) of cassava with differences in cutting lengths, bud
 156 removal treatments and varieties at 360 days after planting (DAP) in 2010 and 2011.

	2010					2011				
	FRY (Mg/ha)	DRY (Mg/ha)	SY (Mg/ha)	SC (%)	HI	FRY (Mg/ha)	DRY (Mg/ha)	SY (Mg/ha)	SC (%)	HI
<i>Cutting length</i>										
15 cm	88.4 ^a	43.3 ^a	22.6 ^a	25.6 ^a	0.54 ^a	74.0 ^a	38.2 ^a	18.5 ^a	25.1 ^a	0.52 ^a
30 cm	75.8 ^b	32.4 ^b	18.3 ^b	24.2 ^b	0.51 ^b	61.0 ^b	32.4 ^b	14.4 ^b	23.7 ^b	0.50 ^b
<i>Bud removal</i>										
Cut	89.1 ^a	44.1 ^a	22.5 ^a	25.3 ^a	0.53 ^a	75.2 ^a	38.1 ^a	18.6 ^a	24.8 ^a	0.52 ^a
Not cut	75.0 ^b	37.6 ^b	18.3 ^b	24.5 ^b	0.52 ^b	59.9 ^b	31.5 ^b	14.3 ^b	24.0 ^b	0.50 ^b
<i>Variety</i>										
KU50	91.4 ^a	48.4 ^a	20.1 ^a	22.0 ^b	0.53 ^a	87.4 ^a	44.7 ^a	18.7 ^a	21.5 ^b	0.52 ^a
HB60	70.8 ^c	38.1 ^b	19.1 ^b	27.0 ^a	0.51 ^b	57.1 ^b	31.0 ^b	15.1 ^b	26.5 ^a	0.50 ^b
RY9	82.6 ^b	39.0 ^b	21.2 ^a	25.7 ^a	0.53 ^a	62.0 ^b	32.6 ^b	15.6 ^{ab}	25.2 ^a	0.51 ^{ab}

157 Means of the same category in the same column followed by different letters are significantly
 158 different at 0.05 probability level by LSD.

159

160 Table 2. Leaf area index (LAI) of cassava with differences in cutting lengths, bud removal
 161 treatments and varieties evaluated at 120, 240, 300 and 360 days after planting (DAP)
 162 in 2010 and 2011.

	2010				2011			
	120 DAP	240 DAP	300 DAP	360 DAP	120 DAP	240 DAP	300 DAP	360 DAP
<i>Cutting length</i>								
15 cm	4.03	1.40	1.64	4.51 ^a	3.95	1.32	1.56	4.43 ^a
30 cm	3.93	1.21	1.57	3.76 ^b	3.85	1.13	1.49	3.68 ^b
<i>Bud removal</i>								
Cut	4.10	1.34	1.62	4.24	4.03	1.27	1.54	4.16
Not cut	3.86	1.27	1.59	4.03	3.78	1.19	1.51	3.96
<i>Variety</i>								
KU50	4.25	1.39	2.37 ^a	4.94 ^a	3.64	1.32	2.29 ^a	4.86 ^a
HB60	3.71	1.29	0.81 ^c	3.56 ^c	4.17	1.22	0.74 ^c	3.48 ^c
RY9	3.99	1.23	1.63 ^b	3.91 ^b	3.91	1.16	1.55 ^b	3.84 ^b

163 Means of the same category in the same column followed by different letters are significantly
 164 different at 0.05 probability level by LSD.

165

166 *Leaf area index (LAI)*

167 Cutting lengths were only significantly different ($P \leq 0.05$) for LAI at 360 DAP in 2010 and
 168 2011 (Table 2). The cuttings of 15 cm length had the highest LAI across varieties and bud removal
 169 treatments. Bud removal treatments did not affect LAI across growth stages. Significant
 170 differences ($P \leq 0.05$) among cassava varieties were observed for LAI at 300 and 360 DAP in 2010
 171 and 2011, with KU50 showing the highest LAI at 360 DAP regardless cutting lengths, bud
 172 removal treatments and years.

173
 174 *Shoot dry weight*

175 Shoot dry weight increased with time for all treatments, with cutting of 30 cm length
 176 causing ($P \leq 0.05$) higher shoot dry weight than cutting of 15 cm length at 360 DAP (Table 3). At
 177 360 DAP, the highest shoot dry weight for the cutting 30 cm length was 2,370 and 2,321 g plant⁻¹
 178 in 2010 and 2011, respectively, whereas the highest shoot dry weight for the cutting of 15 cm
 179 length were 2,196 and 2,149 g plant⁻¹. Cutting without bud removal caused higher shoot dry weight
 180 ($P \leq 0.05$) than cutting with bud removal at 120, 240 and 300 (DAP). Significant differences
 181 ($P \leq 0.05$) among cassava varieties were also observed for shoot dry weight at 120, 240, 300 and
 182 360 DAP across years. HB60 had consistently and significantly the highest shoot dry weight across
 183 sampling times followed by KU50 and RY9. The highest values of shoot dry weight (2,291 g plant⁻¹
 184 in 2010 and 2,247 g plant⁻¹ in 2011) were obtained from HB60 at 360 DAP.

185
 186 Table 3. Shoot dry weight per plant (g) of cassava with differences in cutting lengths, bud removal
 187 treatments and varieties evaluated at 120, 240, 300 and 360 days after planting (DAP) in
 188 2010 and 2011.

	2010				2011			
	120 DAP	240 DAP	300 DAP	360 DAP	120 DAP	240 DAP	300 DAP	360 DAP
<i>Cutting length</i>								
15 cm	772 ^b	1,670 ^b	1,819 ^b	2,196 ^b	639 ^b	1,652 ^b	1,811 ^b	2,149 ^b
30 cm	876 ^a	1,876 ^a	2,027 ^a	2,370 ^a	734 ^a	1,858 ^a	2,017 ^a	2,321 ^a
<i>Bud removal</i>								
Cut	789 ^b	1,730 ^b	1,881 ^b	2,259	652 ^b	1,708 ^b	1,871 ^b	2,231
Not cut	859 ^a	1,815 ^a	1,965 ^a	2,307	721 ^a	1,799 ^a	1,957 ^a	2,239
<i>Variety</i>								
KU50	815 ^b	1,791 ^b	1,939 ^b	2,285 ^b	697 ^b	1,783 ^b	1,924 ^b	2,234 ^{ab}
HB60	848 ^a	1,812 ^a	1,966 ^a	2,291 ^a	715 ^a	1,796 ^a	1,960 ^a	2,247 ^a
RY9	808 ^c	1,716 ^c	1,864 ^c	2,272 ^c	648 ^c	1,697 ^c	1,857 ^c	2,224 ^b

189 Means of the same category in the same column followed by different letters are significantly
 190 different at 0.05 probability level by LSD.

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197 Table 4. Storage root dry weight (g) per plant of cassava with differences in cutting lengths, bud
 198 removal treatments and varieties evaluated at 120, 240, 300 and 360 days after planting
 199 (DAP) in 2010 and 2011.

	2010				2011			
	120 DAP	240 DAP	300 DAP	360 DAP	120 DAP	240 DAP	300 DAP	360 DAP
<i>Cutting length</i>								
15 cm	867	1,996 ^a	2,840 ^a	3,333 ^a	446	1,442 ^a	2,139 ^a	2,723 ^a
30 cm	922	1,633 ^b	2,353 ^b	2,838 ^b	475	1,206 ^b	1,855 ^b	2,238 ^b
<i>Bud removal</i>								
Cut	948	1,928 ^a	2,912 ^a	3,407 ^a	468	1,494 ^a	2,440 ^a	2,807 ^a
Not cut	840	1,701 ^b	2,280 ^b	2,764 ^b	454	1,155 ^b	1,555 ^b	2,154 ^b
<i>Variety</i>								
KU50	990	2,485 ^a	3,680 ^a	4,337 ^a	543	1,928 ^a	3,057 ^a	3,574 ^a
HB60	824	1,395 ^c	1,846 ^c	2,097 ^c	423	985 ^c	1,326 ^c	1,703 ^c
RY9	870	1,564 ^b	2,263 ^b	2,822 ^b	416	1,060 ^b	1,611 ^b	2,165 ^b

200 Means of the same category in the same column followed by different letters are significantly
 201 different at 0.05 probability level by LSD.

202
 203 *Storage root dry weight*

204 Significant differences ($P \leq 0.05$) between cutting lengths were observed for storage root dry
 205 weight at 240, 300 and 360 DAP across years (Table 4), with the highest values being found in
 206 cuttings of 15 cm length. Cutting with bud removal had higher ($P \leq 0.05$) storage root dry weight
 207 than cutting without bud removal at 240, 300 and 360 DAP across years. At final harvest (360
 208 DAP), cutting with bud removal had the highest storage root dry weight of 3,407 and 2,807 g plant⁻¹
 209 in 2010 and 2011, respectively. Cassava varieties were significant different ($P \leq 0.05$) for storage
 210 root dry weight at 240, 300 and 360 DAP across years. KU50 had the highest storage root dry
 211 weight at 240, 300 and 360 DAP in both years. At 360 DAP, KU50 had the highest storage root
 212 dry weight of 4,337 and 3,574 g plant⁻¹ in 2010 and 2011, respectively.

213
 214 *Storage root number per plant*

215 Significant differences ($P \leq 0.05$) between cutting lengths were observed for storage root
 216 number per plant at 120, 240, 300 and 360 DAP in both years (Table 5). Cutting of 15 cm length
 217 caused higher storage root number per plant than that of 30 cm length. Bud removal treatments
 218 were significantly different ($P \leq 0.05$) for storage root number per plant across sampling times, and
 219 cutting with bud removal had higher storage root number per plant than cutting without bud
 220 removal. Cutting with bud removal also had the highest numbers of storage roots at 360 DAP (14.9
 221 and 13.5 roots in 2010 and 2011, respectively). Cassava varieties were significantly different
 222 ($P \leq 0.05$) for storage root number per plant for most sampling times and across years. At 360 DAP,
 223 KU50 had the highest storage root number per plant (14.7 roots in 2010 and 13.7 roots in 2011).

224
 225
 226
 227

228 Table 5. Storage root number per plant of cassava with differences in cutting lengths, bud removal
 229 treatments and varieties evaluated at 120, 240, 300 and 360 days after planting (DAP) in
 230 2010 and 2011.

	2010				2011			
	120 DAP	240 DAP	300 DAP	360 DAP	120 DAP	240 DAP	300 DAP	360 DAP
<i>Cutting length</i>								
15 cm	8.6 ^a	11.7 ^a	13.7 ^a	14.6 ^a	6.3 ^a	10.2 ^a	12.3 ^a	13.2 ^a
30 cm	8.3 ^b	10.9 ^b	12.8 ^b	13.8 ^b	5.4 ^b	9.4 ^b	11.3 ^b	12.4 ^b
<i>Bud removal</i>								
Cut	9.2 ^a	11.9 ^a	13.9 ^a	14.9 ^a	6.5 ^a	10.5 ^a	12.4 ^a	13.5 ^a
Not cut	7.7 ^b	10.7 ^b	12.6 ^b	13.6 ^b	5.2 ^b	9.1 ^b	11.2 ^b	12.1 ^b
<i>Variety</i>								
KU50	9.4 ^a	12.0	13.8 ^a	14.7 ^a	6.7 ^a	10.6 ^a	12.7 ^a	13.7 ^a
HB60	7.6 ^c	10.7	12.7 ^c	13.7 ^c	5.0 ^c	9.0 ^c	11.0 ^c	11.9 ^c
RY9	8.5 ^b	11.2	13.2 ^{ab}	14.2 ^b	5.8 ^b	9.8 ^b	11.8 ^b	12.8 ^b

231 Means of the same category in the same column followed by different letters are significantly
 232 different at 0.05 probability level by LSD.

234 DISCUSSION

235
 236 Sustainable improvements in crop yield and quality are always the main purpose of crop
 237 production. This can be achieved by modifying the genetics of crop plants as well as altering
 238 agronomic practices favoring optimum crop productivity. This research focused on altering
 239 agronomic practice by removing buds of cassava cuttings and adjusting cutting length to increase
 240 storage roots number and ultimately increasing economic root yields. The assumptions of this
 241 study are that the increase in root yield is proportional to the increase in root number and this yield
 242 component can be modified by bud removal. In addition, long cuttings should have higher root
 243 number than short cuttings and cassava genotypes may respond differently to bud removal. As the
 244 practice of bud removal is not common for cassava production, this research should be beneficial
 245 to cassava growers especially the small cassava growers.

247 *Bud removal*

248 Cutting with bud removal significantly increased fresh storage root yield and dry storage
 249 root yield (Table 1) and these results confirmed the claim of cassava growers in Thailand. These
 250 gains were associated with storage root dry weight per plant and storage root number per plant
 251 (Tables 4 and 5). As other studies on bud removal experiment are not available, the direct
 252 comparison of the results with others is not possible. Hypothetically, fibrous roots arise from the
 253 basal cut surface of the cuttings and occasionally from the buds. Some of these fibrous roots start
 254 to bulk and became storage roots (Knoth, 1993). Cutting with bud removal may produce a higher
 255 number of fibrous roots than those of non-cutting bud treatment. This is probably due to the
 256 accumulation of assimilates transported from other parts of the cuttings at the wounds created by
 257 bud removal. The wounds may develop more fibrous roots and increase the possibility of these
 258 roots to become storage roots.

259 In a classical experiment conducted in 1686, phloem tissue was cut by removing the bark.
260 The assimilates produced in leaves were transported along the phloem tissue and stopped at the
261 wound, resulting in high accumulation of assimilates at the wound and the development of roots
262 (Malpighi, 1686). Wound-induced roots have been reported in geranium (Davies *et al.*, 1982; Cline
263 and Neely, 1983) and woody plant (Jackson, 1986). This knowledge is commonly used for plant
264 propagation such as cutting and layering. Stem cuttings with removal of buds, bark (phloem tissue)
265 and cambium layer at underground level increased root number and also increased the possibility
266 of these roots (fibrous roots) to become storage roots. As consequences, bud-removal treatment
267 caused higher fresh storage root yield, dry storage root yield, starch yield, starch content, harvest
268 index, storage root dry weight and storage root number per plant (Tables 1, 4 and 5). However, the
269 experiment focused on storage roots and aerial growth and the formation of fibrous roots at early
270 growth stages was not investigated. Further deliberate experiments are required to verify the
271 hypothesis for yield increase as affected by bud removal.

272

273 *Cutting length*

274 The question underlying this research is what length is suitable for bud removal practice and
275 then cuttings of 15 and 30 cm length were compared as these lengths are commonly used for
276 cassava production. In this study, cutting of 15 cm caused higher storage root yield than that of 30
277 cm and this is partially explained by higher LAI, storage root dry weight per plant and storage root
278 number (Tables 2, 4 and 5). The results were in agreement with those of Tongglum *et al.* (1992),
279 who found that cutting of 20 cm had higher storage root yield than that of 25 cm. Another work
280 also reported the advantage of short cuttings over long cuttings (Villamayor *et al.*, 1992). Ganado
281 (1956) found that short cuttings were better than long cuttings when the cuttings were planted
282 vertically, and long cuttings were better than short cuttings when the cuttings were planted
283 horizontally. However, cuttings of 10 and 50 cm were not significantly different for root yield
284 (Velasco, 1982). Long cuttings with more than 10 nodes had a better chance of conserving their
285 viability, and stem cuttings of 5 to 7 nodes and minimum length of 20 cm were recommended to
286 obtain optimum yield (Carvalho *et al.*, 1993). Cuttings with 4 to 7 nodes were not different with
287 respect to mean storage root length, radius of storage root tip and the number of major stem per
288 plant (Onwueme, 1978). However, longer cuttings produced a faster growing canopy (Lahai *et al.*,
289 1999). The differences among above studies would be due to differences in planting methods
290 (vertical insert and horizontal insert) and environments (tillage and soil moisture).

291 In this study, cutting of 30 cm had higher shoot dry weight per plant than that of 15 cm,
292 which was associated with higher stem dry weight (data not shown). However, higher shoot dry
293 weight of cutting of 30 cm length did not cause root yield advantage and this would be due to
294 lower starch content, harvest index and root number (Tables 1, 3 and 5). The assumption is that a
295 longer cutting has higher sprouting and development due to the presence of more buds. Storage
296 root number and root yield were shown to be affected by cutting length and root yield was
297 associated with the number of storage roots (Didier and El-Sharkawy, 1994). The higher LAI
298 associated with cutting of 15 cm indicates that plants can produce higher levels of photosynthate,
299 which has been correlated positively with root yield (Lenis *et al.*, 2005; Lebot, 2009; Lahai *et al.*,
300 2013). During the period of 120 to 180 DAP, plants maintained LAI above 4.0 (2010) and 3.9
301 (2011), with LAI subsequently declining to 1.4 in 2010 and 1.3 in 2011 for the remainder of their
302 growth cycle. This pattern is due to leaf senescence and abscission during rainless period (Figure
303 1). Following the onset of rainy season, new leaves were produced and LAI values above 4.5
304 (2010) and 4.4 (2011) were recorded (Table 2; Figure 1).

305 In the present study, the long cutting (30 cm) had higher shoot dry weight (Table 3) due to
306 the presence of higher reserved carbohydrate. However, lower LAI in long cutting might be
307 attributed to the limitation of nutrients applied to the soil. The application of chemical fertilizer
308 formula 15-15-15 (N₂, P₂O₅, K₂O) at the rate of 312.5 kg ha⁻¹ is generally recommended for
309 cuttings of 15 to 20 cm. Long cutting produced higher branch number, but it had lower LAI
310 possibly due to poor partitioning of assimilates to the branches. Although long cutting produced
311 higher number of fibrous roots, the formation of storage roots was low due to low LAI (Table 2).
312 Shoots depend on roots for nutrient and water uptake, while the continued root growth is reliant
313 on photosynthates produced by leaves (Kramer and Boyer, 1995). Crop performance in 2011 was
314 generally lower than in 2010 for most parameters investigated. The reduction in crop performance
315 was in large part due to rainfall amount and distribution during the growing season. The crop
316 received a total rainfall of 1,621 mm in 2011 and 1,099 mm in 2010. Then, water logging occurred
317 during high rainfall intensity in September 2011, and resulted in the reduction in crop growth and
318 yield as compared to 2010 (Figure 1).

319

320 *Cassava variety*

321 KU50 had the highest fresh storage root yield and dry storage root yield in 2010 and 2011,
322 which was associated with the highest LAI, storage root dry weight per plant and storage root
323 number (Tables 1, 2, 4 and 5). In previous investigations, high LAI is an important factor leading
324 to high yield in cassava varieties (Enyi, 1973; Lahai, 1999). On the other hand, HB60 had the
325 highest starch content in storage root. Accordingly, Vichukit *et al.* (2004) reported that HB60 had
326 higher starch content than RY5, RY72 and KU50. Herein, KU50 produced the highest starch yield
327 due to the highest fresh storage root yield. KU50 also had higher HI than did HB60, indicating that
328 KU50 was highly efficient in transporting photoassimilates for storage in tuber roots. KU50 is
329 popular among cassava growers in the Northeast, Thailand, being widely adapted to unfavorable
330 growing conditions (Rojanaridpiched *et al.*, 1995). In this study, only three cassava varieties were
331 investigated to understand the responses to bud removal and cutting lengths. The results indicated
332 that the varieties responded similarly in terms of rooting and formation of storage roots. However,
333 more cassava varieties should be investigated to reach a recommendation of management practice
334 to cassava growers. In addition, the experiment was conducted in the early rainy season planting
335 date and results do not cover the planting date in the late rainy season. This is the main planting
336 date for cassava growing in Thailand and such seasonal influence on cassava management should
337 be further investigated.

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CONCLUSIONS

340 Bud removal practice did increase cassava yield in all evaluated varieties and the
341 combination for the best yield was bud removal with cutting of 15 cm. The interaction between
342 cutting length and environment should be considered. KU50 had the highest fresh storage root
343 yields.

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REFERENCES

- 359 Alves A. A. C. (2002). Cassava botany and physiology. In *Cassava: Biology, production and*
360 *utilization*, 67-90 (Eds R. J. Hillocks, J.M. Thresh and A. Bellotti). CABI, Wallingford, UK.
- 361 Bainbridge, Z., Tomlins, K., Wellings, K. & Westby, A. (1996). *Methods for Assessing Quality*
362 *Characteristics of Non-Grain Starch Staples*. (Part 2. Field Methods.). Natural Resources
363 Institute, Chatham, UK.
- 364 Bouyoucos, G. J. (1962). Hydrometer method improved for making particle size analysis of soils.
365 *Agronomy Journal* 54:464-465.
- 366 Bray, R. H. & Kurtz, L. T. (1945). Determination of total, organic and available forms of
367 phosphorus in soils. *Soil Science* 59:39-45.
- 368 Bremner, J. M. (1965). Total nitrogen. In *Methods of Soil Analysis. Part 2: Chemical and*
369 *Microbiological Properties*, 1149–1178 (Ed C. A. Black). Madison, Wisconsin, USA.
- 370 Carvalho, L.J., Mattos-Cascardo, C.B.J., Ferreria, M. & Loureiro, M. (1993). Studies on proteins
371 and enzymes related to tuberization and starch biosynthesis in cassava roots. In *International*
372 *Scientific Meeting of the Cassava Biotechnology Network*, 234-238 (Eds W. Roca and A.
373 Thro). Proc. 1st, held in Cali, Columbia.
- 374 Cline, M. N. & Neely, D. (1983). The histology and histochemistry of the wound healing
375 process in geranium cuttings. *Journal of the American Society for Horticultural Science*
376 108:450-96.
- 377 Davies, F. T., Lazarte, J. E. & Joiner, J. N. (1982). Initiation and development of roots in
378 juvenile and mature leaf bud cuttings of *Ficus pumila* L. *American Journal of Botany*
379 69:804-811.
- 380 Didier, P. & El-Sharkawy, M. A. (1994). Sink-source relations in cassava effects of reciprocal
381 grafting on yield and leaf photosynthesis. *Experimental Agriculture* 30:359-367.
- 382 Ekanayake, I. J. (1994). Terminology for growth analysis of cassava. *Tropical Root and Tuber*
383 *Crops Bulletin* 8:2-3.
- 384 Enyi, B. A. C. (1973). Growth rates of three cassava varieties (*Manihot esculenta* Crantz) under
385 varying population densities. *The Journal of Agricultural Science* 81:15-28.
- 386 FAO. 2014. Food and Agriculture Organization. Corporate Document Repository. Available at:
387 <http://faostat.fao.org/site/339/default.aspx>., verified 19 November, 2014.
- 388 FAO. 2014. Food and Agriculture Organization. FAO 2014 Database. Available at:
389 <http://www.fao.org/docrep/007/y2413e/y2413e0i.htm>., verified 20 November, 2014.
- 390 Ganado, Z. S. (1956). *The effect of length of cassava cutting on yield*. B.Sc. Thesis, CPU, Iloilo
391 City.
- 392 Hahn, S. K. & Hozyo, Y. (1984). Sweet potato. In *The Physiology of Tropical Field Crops*, 551-
393 576 (Eds P. R. Goldsworthy and N. M. Fisher). John Wiley and Sons, New York.
- 394 Jackson, M. B., ed. (1986). *New root formation in plants and cuttings*. Dordrecht: Martinus
395 Nijhoff Publishers.

396 Jones, S., Robin, G., Garry, J., Johnson, L. & Benn, A. (2013). Production, productivity, quality
397 standards and product mixes of roots and tubers crops in the CFC-funded project. Publication
398 DO/029/12. Available at: [http://www.cardi.org/cfc-rt/files/downloads/2013/09/Publ-9-](http://www.cardi.org/cfc-rt/files/downloads/2013/09/Publ-9-Production-Productivity-RT-Jones-et-al.pdf)
399 [Production-Productivity-RT-Jones-et-al.pdf](http://www.cardi.org/cfc-rt/files/downloads/2013/09/Publ-9-Production-Productivity-RT-Jones-et-al.pdf). verified 19 February, 2016.

400 Knoth, J. (1993). Traditional storage of yams and cassava and its improvement. Deutsche
401 Gesellschaft für Technische Zusammenarbeit (GTZ) GmbH, Postfach 5180, D-65726
402 Federal Republic of Germany. Available at:
403 [http://www.fastonline.org/CD3WD_40/INPHO/VLIBRARY/GTZHTML/X0066E/EN/X0](http://www.fastonline.org/CD3WD_40/INPHO/VLIBRARY/GTZHTML/X0066E/EN/X0066E00.HTM)
404 [066E00.HTM](http://www.fastonline.org/CD3WD_40/INPHO/VLIBRARY/GTZHTML/X0066E/EN/X0066E00.HTM), verified 19 February, 2016.

405 Knutsson, J. (2012). Long-term Storage of Starch Potato and its Effect on Starch Yield. M.Sc.
406 thesis. Swedish University of Agricultural Sciences. Available at:
407 http://stud.epsilon.slu.se/4996/1/knutsson_j_121023.pdf, verified 19 February, 2016.

408 Kramer, P. J. & Boyer, J. S. (1995). Roots and root systems. In *Water Relations of Plants and Soil*,
409 115-166 (Eds P. J. Kramer and J. S. Boyer). San Diego, California.

410 Lahai, M. T., George, J. B. & Ekanayake, I. J. (1999). Cassava (*Manihot esculenta* Crantz) growth
411 indices, root yield and its components in upland and inland valley ecologies of Sierra Leone.
412 *Journal of Agronomy and Crop Science* 182:239-248.

413 Lahai, M. T., Ekanayake, I. J. & Koroma, J. P. C. (2013). Influence of canopy structure on yield
414 of cassava cultivars at various toposequences of an inland valley agro ecosystem. *Journal of*
415 *Agricultural Biotechnology and Sustainable Development* 5:36-47.

416 Lebot, V. (2009). *Tropical root and tuber crops: cassava, sweet potato, yam and aroids*. CABI
417 publication. Amazon.com.

418 Leihner, D. (2012). Agronomy and Cropping Systems. In *Cassava: Biology, Production and*
419 *Utilization*, 91–113 (Eds R. J. Hillocks, J. M. Thresh and A. C. Bellotti). CAB International,
420 Wallingford, UK.

421 Lenis, J. I., Cali, F., Jaramillo, G., Perez, J. C., Ceballos, H. & Cock, J. H. (2005). Leaf retention
422 and cassava productivity. *Field Crops Research* 95:126-134.

423 Malpighi, M. (1686). *Opera Omnia*. Royal Society of London, London.

424 McKeague, J. A. (1978). *Manual on soil sampling and methods of analysis-2nd Edition*. (Canadian
425 Society of Soil Science, Suite 907, 151 Slater St., Ottawa, Canada).

426 Nassar, N. M. A. & Teixeira, R. P. (1983). Seed germination of wild cassava species (*Manihot*
427 *spp.*). *Ciencia e Cultura* 35:630-632.

428 OAE. (2014). Office of Agricultural Economics. Thai Economics Database 2014. Available at:
429 <http://www.oae.go.th/download/prcai/DryCrop/amphoe/casava-amphoe57.pdf>, verified
430 16 November, 2014.

431 Onwueme, I. C. (1978). *The Tropical Tuber Crops: Yam, Cassava, Sweet Potato and Cocoyam*.
432 John Wiley and Sons, New York.

433 Osiru, D. S. O., Porto, M. C. M. & Ekanayake, I. J. (1997). *Physiology of Cassava*. IITA Research
434 Guide 55. Training Program, IITA, Ibadan, Nigeria.

435 Rojanaridpiched, C., Limsila, A., Supraharn, D., Boonseng, O., Poolsanguan, P., Tiraporn, C. &
436 Kawano, K. (1995). Recent progress in cassava varietal improvement in Thailand. In
437 *Cassava Breeding, Agronomy Research and Technology Transfer in Asia*, 124-134 (Ed R.
438 H. Howeler). Proc. 4th Regional Workshop, held in Trivandrum, Kerala, India.

439 Scott, G. J., Rosegrant, M. W. & Ringler, M. W. (2000). Roots and Tubers for the 21st Century:
440 trends, projections and policy options. Food, Agriculture and the environment discussion

441 paper 31. International Food Policy Research Institute. Available at:
442 http://pdf.usaid.gov/pdf_docs/Pnach747.pdf, verified 19 February, 2016.

443 Tongglum, A., Vichukit, V., Jantawat, S., Sittibusaya, C., Tiraporn, C., Sinthuprama, S. &
444 Howeler, R. H. (1992). Recent progress in cassava agronomy research in Thailand. In
445 *Cassava Breeding, Agronomy and Utilization Research in Asia*, 199-223 (Ed R. H.
446 Howeler). Proc. 3rd Regional Workshop, held in Malang, Indonesia. Oct 22-27, 1990.

447 Velasco, O. L. (1982). The effects of different lengths of cutting on the growth and yield of
448 cassava. B.Sc. thesis. WLAC, San Marcelino, Zambales.

449 Villamayor, F. G., Dingal, A. G., Evangelio, F. A., Ladera, J. C., Medellin, A. C., Sajise, G. E. &
450 Burgos, G. B. (1992). Recent progress in cassava agronomy research in the Philippines. In
451 *Cassava Breeding, Agronomy and Utilization Research in Asia*, 245-259 (Ed R. H. Howeler
452) Proc. 3rd Regional Workshop, held in Malang, Indonesia.

453 Vichukit, V., Rodjanaridpiched, C., Poonsaguan, P., Sarobol, E., Cheamchamnuncha, C.,
454 Changlek, P., Piyachomkwan, K. & Sriroth, K. (2004). Huay Bong 60: New developed Thai
455 cassava (*Manihot esculenta* Crantz) variety with improved starch yield and quality. In *The*
456 *6th International Scientific Meeting of the Cassava Biotechnology Network (CBN)*. March 8-
457 14, 2004. Cali, Colombia.

458 Walkley, A. & Black, I. A. (1934). An examination of Degtjareff method for determining soil
459 organic matter and a proposed modification of the chromic acid titration method. *Soil*
460 *Science* 37:29-37.

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464 Supplementary material

465 **Table S1.** Baseline soil physical and chemical properties prior to planting in 2010 and 2011

Soil properties	2010	2011
<i>Chemical</i>		
pH ¹	6.1	6.2
Total nitrogen (%) ²	0.034	0.015
Available phosphorus (Mg kg ⁻¹) ³	21.7	9.3
Exchangeable potassium (Mg kg ⁻¹) ⁴	32.6	16.7
Organic matter (%) ⁵	0.58	0.34
<i>Soil texture⁶</i>		
Sand (%)	78.0	84.9
Silt (%)	13.0	8.9
Clay (%)	9.0	6.1
Texture	loamy sand	loamy sand

466 ¹ = pH method

467 ² = Micro-Kjedahl method (Bremner, 1965)

468 ³ = Bray II method (Bray and Kurtz, 1945)

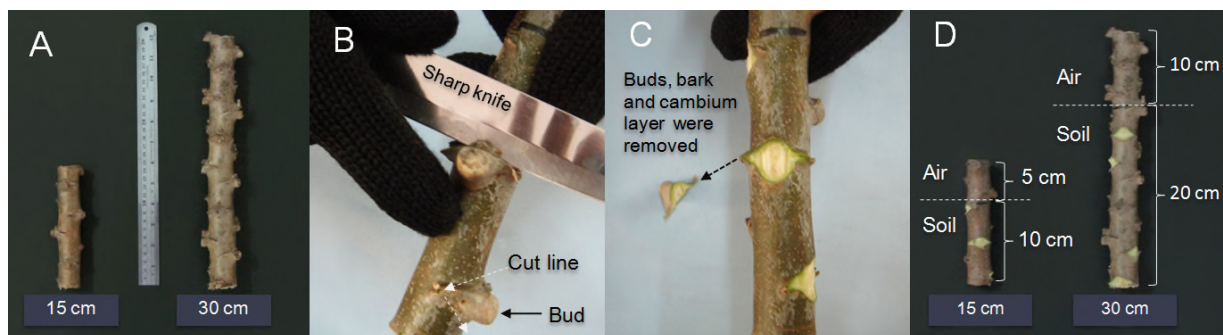
469 ⁴ = 1 N ammo-nium acetate pH =7.0 (McKeague, 1978)

470 ⁵ = Walkley and Black (Walkley and Black, 1934)

471 ⁶ = Hydrometer method (Bouyoucos, 1962)

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476 **Figure S1.** The process of removing buds from stem cuttings: A, cuttings of 15 and 30 cm lengths,
 477 B, buds at underground level being cut; C, buds at underground level after cut; and D, cuttings of
 478 15 and 30 cm length showing underground part and aerial part.