

Full length article

Severity scoring of manganese health effects for categorical regression



Donald R. Mattison^{a,b,*}, Brittany Milton^a, Daniel Krewski^{a,b}, Len Levy^c,
David C. Dorman^d, Peter J. Aggett^e, Harry A. Roels^f, Melvin E. Andersen^g,
Nataliya A. Karyakina^{a,b}, Natalia Shilnikova^{a,b}, Siva Ramoju^a, Doreen McGough^h

^a Risk Sciences International, 55 Metcalfe Street, Suite 700, K1P 6L5, Ottawa, Canada

^b R. Samuel McLaughlin Centre for Population Health Risk Assessment, Faculty of Medicine, University of Ottawa, 118-850 Peter Morand Drive, Canada

^c Institute of Environment and Health, Cranfield University, College Road, Cranfield MK43 0AL, Bedfordshire, United Kingdom

^d College of Veterinary Medicine, North Carolina State University, 1060 William Moore Drive, Raleigh, NC 27607, USA

^e School of Medicine and Health, Lancaster University, Bailrigg, Lancaster, LA1 4YW, United Kingdom

^f Louvain Centre for Toxicology and Applied Pharmacology (LTAP), Université catholique de Louvain, Avenue Mounier 53.02, 1200 Brussels, Belgium

^g ScitoVation, 6 Davis Drive, PO Box 110566, Research Triangle Park, NC, 27709-2137, USA

^h International Manganese Institute, 17 rue Duphot, 75001 Paris, France

ARTICLE INFO

Article history:

Received 21 July 2016

Received in revised form 24 August 2016

Accepted 4 September 2016

Available online 13 September 2016

Keywords:

Exposure-response assessment

Categorical regression

Database

Manganese toxicity

ABSTRACT

Characterizing the U-shaped exposure response relationship for manganese (Mn) is necessary for estimating the risk of adverse health from Mn toxicity due to excess or deficiency. Categorical regression has emerged as a powerful tool for exposure-response analysis because of its ability to synthesize relevant information across multiple studies and species into a single integrated analysis of all relevant data. This paper documents the development of a database on Mn toxicity designed to support the application of categorical regression techniques. Specifically, we describe (i) the conduct of a systematic search of the literature on Mn toxicity to gather data appropriate for dose-response assessment; (ii) the establishment of inclusion/exclusion criteria for data to be included in the categorical regression modeling database; (iii) the development of a categorical severity scoring matrix for Mn health effects to permit the inclusion of diverse health outcomes in a single categorical regression analysis using the severity score as the outcome variable; and (iv) the convening of an international expert panel to both review the severity scoring matrix and assign severity scores to health outcomes observed in studies (including case reports, epidemiological investigations, and in vivo experimental studies) selected for inclusion in the categorical regression database. Exposure information including route, concentration, duration, health endpoint(s), and characteristics of the exposed population was abstracted from included studies and stored in a computerized manganese database (*MnDB*), providing a comprehensive repository of exposure-response information with the ability to support categorical regression modeling of oral exposure data.

© 2016 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

Manganese (Mn) is a naturally occurring element and an essential nutrient. Dietary intake of Mn is essential for maintaining a number of important physiological processes, including reproduction and development (e.g., formation of healthy cartilage and

bone), energy metabolism (e.g., pyruvate carboxylase), urea cycle (e.g., arginase), and antioxidative capacity (e.g., Mn superoxide dismutase) (Chen et al., 2014). Mn also plays a key role in wound-healing (ATSDR, 2012). Mn is found in nutritional supplements and multivitamin preparations (Santos-Burgoa et al., 2001).

There is a large body of scientific literature on adverse health effects associated with excess or deficient levels of Mn. The toxicity of Mn due to excess or deficiency has been documented in diverse studies including case reports, epidemiological studies of occupational and environmental exposure to Mn, experimental studies in a range of animal models, and in vitro toxicity tests. Krewski et al. (2010) describe an approach to incorporation of data from a diverse

* Corresponding author at: Risk Sciences International, 55 Metcalfe Street, Ottawa K1P 6L5, Canada.

E-mail addresses: dmattison@risksciences.com, mattisond@aol.com (D.R. Mattison), doreen.mcough@manganese.org (D. McGough).

collection of studies of this nature based on categorical regression of severity scores assigned to the different health outcomes seen in these studies; the utility of this approach was demonstrated by application to a database on copper toxicity, similar to the manganese database (*MnDB*) developed here. This copper database (*CuDB*) was subsequently analyzed by Chambers et al. (2010) to describe the U-shaped exposure response curve for Cu, which, like Mn, is an essential element. Further analyses of the *CuDB* were recently undertaken by Milton et al. (2016a), where they employed new approaches to categorical regression analysis of U-shaped exposure-response curves. In conducting this work, the available data on Cu toxicity due to both excess and deficiency was entered into a computerized database designed to accommodate the collection of information on continuous, dichotomous, categorical or ordinal data which supports both traditional as well as new methods for exposure-response assessment. Further motivation for the use of a systematic approach to the identification and recording of relevant data on Cu toxicity is to avoid unnecessary repetition of reviews of the same literature: without a validated toxicological data storage system, changing regulatory requirements, updating risk assessments, and employing new methods for exposure-response assessment would likely involve unnecessary re-reviews of the same body of literature (Guth and Raymond, 1996).

In the field of health risk assessment, the characterization of exposure-response relationships is important in estimating the risk of adverse health effects of essential elements from toxicity due to either excess or deficiency. Health risk scientists have not yet defined exposure-response curves that simultaneously characterize the risk associated with both Mn deficiency and excess. Historically, regulatory agencies have used benchmarks such as the no-observed-adverse-effects level (NOAEL), corresponding to the level of exposure that does not result in a significant increase in the risk of adverse effects in the exposed group when compared with controls: the NOAEL has served as a point of departure (PoD) on the exposure response curve for establishing a reference dose (RfD) for human exposure through the application of appropriate adjustment factors (Barnes and Dourson, 1988). These benchmarks are typically derived from a single key study that considers one critical effect and rely on weight of evidence assessment for relevant effect in humans and to a considerable extent on expert opinion. This led to differences in human exposure guidelines developed by different regulatory bodies (US EPA, 1993, 1994; Health Canada, 1994; ATSDR, 2000, 2012; WHO, 2000), including occupational exposure guidelines (Deveau et al., 2015). This is illustrated by the disparity of health-based limit values for inhalation of respirable Mn particulate in ambient air (ranging from 0.04 to 0.30 $\mu\text{g}/\text{m}^3$) derived from the same epidemiological study of battery workers exposed to MnO₂ dust (Roels et al., 1992).

More recently, exposure-response assessment methods have shifted towards more quantitative methods, with health risk assessors exploring more mathematically driven techniques such as the benchmark dose (BMD) (Crump, 1984), and signal-to-noise crossover dose (SNCD) (Sand et al., 2011). Nonetheless, the RfD, SNCD, and BMD approaches all ultimately rely on one critical health effect from a single key study.

Categorical regression addresses this limitation by allowing risk assessors to capture relevant health information across multiple studies and species, including a broad spectrum of health endpoints and exposure levels for exposure-response analysis in an objective and transparent manner. Furthermore, categorical regression also allows the inclusion of multiple independent variables, including level and duration of exposure, and variables that may modify the exposure-response relationship such as age and sex. For these reasons, categorical regression has been advocated as a promising tool to characterize health risk in a

comprehensive manner, and has found successful initial application in exposure-response modeling (Gift et al., 2008; Allen et al., 2005; Chambers et al., 2010).

Ten years ago, the US EPA (2006) released a software program called *CatReg*, developed to perform categorical regression modeling and calculate a benchmark level called the extra risk concentration (ERC). Chambers et al. (2010) used *CatReg* to perform an exposure-response analysis on the copper database previously described, creating separate excess and deficiency exposure-response models for oral intake. The authors spliced the excess and deficiency curves together to create a U-Shaped curve, then estimated the exposure level at the trough of the curve. Other *CatReg* applications include hydrogen sulfide (Strickland and Foureman, 2002; Brown and Strickland, 2003; Brown and Foureman, 2005), phosgene (Gift et al., 2008), and acrylamide (Allen et al., 2005), where excess exposure-toxicity curves were fit to exposure-response data. Milton et al. (2016a) used the work by Chambers et al. (2010) as a platform to propose a new method for defining U-Shaped exposure-response curves based on categorical regression. The authors applied their methods to the copper (Cu) toxicity database and obtained a smooth, continuous U-Shaped exposure-response curve that achieves balance between Cu excess and deficiency. The authors identified two potential benchmark levels: the equiprobable crossover point (EPCP), which corresponds to the level of exposure where the risk of toxicity due to excess is equal to the risk of toxicity due to deficiency, and X_{MINDUE} , which corresponds to the level of exposure at the bottom of the U-shaped which minimizes the overall risk due to excess or deficiency (or both). The methodologies used to derive this U-shaped exposure-response curve and the estimation of these two new benchmarks for Mn are discussed in a companion paper (Milton et al., 2016b).

These new approaches to categorical regression modeling developed by Milton et al. (2016a) will be used in the manganese exposure-response assessment. The foundation of categorical regression modeling is the establishment of ordered response categories corresponding to increasingly severe adverse health outcomes and the availability of a comprehensive database which summarizes ordered response categories for manganese toxicity from deficiency or excess.

The purpose of this paper is to: 1) describe the development of the computerized Mn database (*MnDB*) to support the application of categorical regression of Mn toxicity due to excess and deficiency from oral studies; 2) to summarize the development of the severity scoring system for Mn toxicity; 3) to apply the severity scoring system to the scientific literature collected on Mn health effects; and 4) describe the characteristics of the final *MnDB* and its use in categorical regression (Fig. 1).

2. Methods

The development of the categorical regression database took place over the course of two years (2010–2012). Exclusion criteria were defined and relevant scientific publications were identified using a systematic literature search and reviewed to ensure the exclusion criteria were satisfied. A total of 181 eligible studies described in 218 articles were identified (Appendix A). Detailed information including animal species, route of exposure, Mn species, age, sex, study design, dose and duration of exposure, and health outcome was abstracted from these articles and stored in the database. If a study involved different exposure scenarios (e.g., different exposure routes and pathways, different doses and concentrations, different exposure durations, different Mn compounds and basal diets, animal species and strains, sex (male and female subjects)), data for each combination of these parameters were entered as a separate experiment. In total, the present version

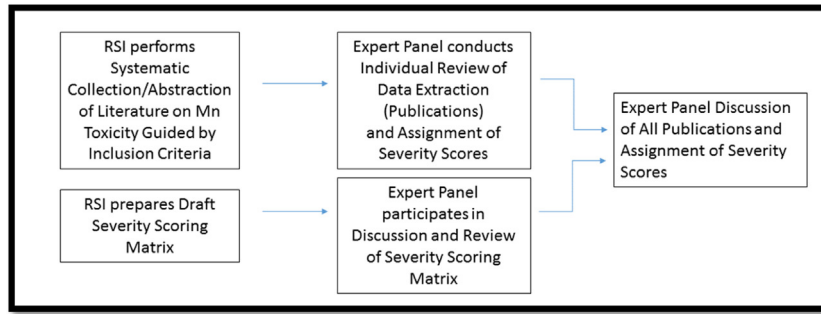


Fig. 1. Work flow diagram of the development and application of the severity scoring matrix to the MnDB.

of the *MnDB* includes data from 272 experiments. There are generally several dose levels within a single experiment, with a separate record created for each dose level. Some studies are described in more than one article: in this event, information from these articles was combined so as to avoid duplication in the *MnDB*.

Upon completion of the *MnDB*, a draft ordinal severity scoring matrix covering the spectrum of health outcomes in the *MnDB* was created. A three-day workshop was held at Risk Sciences International in Ottawa, Canada at the end of January 2013 with participation of experts in epidemiology, toxicology, medicine, veterinary sciences, and risk science. The expert panel was charged to review and modify the ordinal scale of severity scores and apply it to the health outcomes in the computerized database. The expert panel also modified and endorsed the study exclusion criteria specified below in Section 2.1.

2.1. Literature search and exclusion criteria

To develop a robust categorical regression database, it was important to first identify relevant scientific reports for inclusion in the database. To achieve this, the International Manganese Institute (IMNI) electronic library reference list as well as Ovid Medline/Embase and Toxline bibliographic databases were searched. Search terms are provided in Appendix B. No limits were applied to publication date; studies published as early as 1930 and as late as 2013 were included in the analysis. References of identified articles were also searched to identify further relevant publications. Case reports, epidemiological studies and in vivo experimental studies were considered as potentially eligible for inclusion.

The international expert panel also guided the modification of exclusion criteria. For example, it was suggested that studies with transgenic animals with altered metabolic profiles that might be of limited relevance to human health risk assessment be excluded from database until scientific data (e.g., PBPK modeling data) is available to compare dose metrics against conventional animal models. It was also suggested that these studies be retained as a separate group in the *MnDB* for possible use in categorical regression sensitivity analysis. Similarly, arguments for inclusion of metabolic/pharmacokinetic and in vitro studies could be made, as these studies may be useful in elaborating toxicity pathways for Mn. For example, in experiments in which neurotransmitters were evaluated, in vitro studies might be useful in determining severity level of the potential adverse outcomes. However, in the absence of formal criteria for incorporating information from pharmacokinetic and in vitro studies into the assignment of severity scores to support categorical regression, the use of such data was not considered in the present exercise.

The final exclusion criteria reflect the modifications and suggestions provided by the expert panel. The exclusion criteria are:

- inadequate information to characterize the dose and/or duration of exposure;
 - the information could not be entirely attributed to the effects of manganese alone (due to the presence of possible confounding);
 - the exposure route was not relevant for humans;
 - exposure occurred in utero;
 - exposure occurred by lactation;
 - the animal model was not considered suitable for human health risk assessment (ruminant species, non-mammals)
 - the study focussed on validation of potential exposure biomarkers (e.g. Mn in blood and urine);
 - there was inadequate statistical reporting of data;
 - the study focused on pharmacokinetic parameters, or Mn body burden;
 - the study was conducted in an in vitro test system (which is difficult to extrapolate to human exposure-response);
 - the article was a review rather than original research study.
- Exclusion criterion (2) was further developed for application to epidemiologic studies, excluding studies with:
- no “external” measures of exposure (e.g. Mn in air), wherein only biomarkers were used as exposure metrics;
 - data on exposure duration were not available;
 - exposure estimates were based on modeling rather than measurement;
 - it was unclear if the measurements of exposure reported total, inhalable or respirable Mn dust.

2.2. Characteristics of the database

The database was created in Microsoft Access and contains a wide collection of variables, ranging from qualitative inputs related to data abstraction/storage to quantitative inputs associated with exposure. The **identifier variable** is an ID automatically assigned to each record. The **identifier variable** also contains the first author's last name, publication year, and the full reference. Note that a common study ID is assigned to all experiments within the same study. Characteristics of study subjects, such as **species, strain, sex, and life stage** at first exposure (e.g. newborn, weanling, adult, aged) appear in the database. Furthermore, the characteristics of exposure, namely, the **manganese compound, exposure route** (oral or inhalation), **exposure medium** (food, drinking water or gavage for oral exposure; dust or fume for inhalation exposure), **dose of Mn** (mg/kg bw/day or concentration of Mn in air ($\mu\text{g}/\text{m}^3$)), and **duration of exposure** in days also appear in the database.

Each outcome under investigation was described in a separate text field. An **ordinal severity score** was assigned to each outcome on the basis of a severity system described in the following section. Because neurotoxic effects are “critical” for Mn health risk assessment, each experiment in the database has an indicator of

- exposure to organic manganese (Mn) compounds;

whether or not this experiment contains at least one neurotoxicity-related outcome. The highest severity score associated with a neurotoxicity-related outcome at each dose level in each experiment was extracted into a separate field. While neurotoxic outcomes are of interest, *CatReg* modeling exercises could consider any and all health outcomes, not only neurotoxicity.

Following complete data abstraction from the scientific publications included in the database, the data was made available to the expert panel for their independent review and assignment of severity scores for each health endpoint measured and included in the database.

2.3. Development of severity scoring template

All relevant animal and human studies on Mn excess and deficiency were identified. Investigators at Risk Sciences International (RSI) with expertise in toxicology, epidemiology, medicine and risk science applied a systematic approach for the examination and differentiation of the reported Mn effects. Using a severity scoring system, these Mn effects were evaluated based on their relevance to humans and the type and magnitude of toxic effects to create a common measure of the physiological and/or pathophysiological response for application across all studies on Mn excess and deficiency. The overall approach for the development of the severity scoring matrix was guided by a similar original scoring exercise for Cu (Krewski et al., 2010; Chambers et al., 2010), with appropriate modifications based on the Mn-specific mechanism of toxicity and target organs. Changes in the Mn toxicokinetic parameters, biochemical and/or cellular changes involved in Mn toxicity pathways, changes in body/organ weight, organ/system impairment or histopathological changes, and reversibility or irreversibility of these changes were used for evaluation of the severity of effect. A severity scoring matrix was created ranging from low to high severity level (from level 0 to level 9 in the excess severity scoring template and from level 0 to level 8 in the deficiency severity scoring template) and was used to rank the severity of all observed effects in animals and humans according to the organ affected and biochemical effects and/or histopathological effects. For example, in the excess severity scoring template, the lower severity level (severity level 0) was associated with exposures with no observed changes compared to controls (effectively the no-observed-adverse-effect level, or NOAEL); severity level 1 corresponded to homeostatic changes in the observed effects of Mn; level 2 was associated with early adaptive systemic changes of unknown clinical significance; level 3 was associated with lowest-observed-adverse-effect level with biochemical and/or cellular changes involved in Mn toxicity pathways (the lowest-observed-adverse-effect-level, or LOAEL); and level 4 reflected a more severe adverse effect level associated with metabolic perturbations. Severity levels 5–9 represented increasingly severe adverse health outcomes. The highest severity levels 7, 8, and 9 were associated with reversible severe clinical signs of toxicity and histopathological changes, irreversible neurotoxic effects and histopathological changes, and death, respectively. The effects observed in animals and humans under conditions of Mn deficiency are different from the effects observed under Mn excess exposure due to a different mechanism of toxicity following inadequate levels of this essential element in the body. The most severe scores for deficiency, –6, –7 and –8 were associated with reversible clinical signs of deficiency and histopathological changes, irreversible histopathological changes and birth defects and death, respectively.

Table 1 presents the 9 severity categories under excess exposure and the 8 severity categories under deficiency exposure and the corresponding adverse health effects associated with each

level of severity. As a result of this exercise, all outcomes reported in each single study were categorized and scored consistently across all severity levels. Experts' opinion was used to revise and refine the adopted approach, scoring matrix, and assigned scores to the endpoints extracted from studies on both Mn deficiency and excess to use in the exposure-response analysis.

The experts highlighted important issues in the consideration and interpretation of the severity of adverse health outcomes. Specifically, the need to distinguish between reversible and non-reversible effects for excess and deficiency, and between adverse and non-adverse observed outcomes was noted. It was also suggested that Mn accumulation in target organs (brain and lungs) versus non-target organs and tissues (blood, kidneys, urine) be considered, and that the applicability of histopathological considerations in case of the histological changes without reported statistical significance be evaluated. Consideration of the observed clinical signs as sufficient evidence of an adverse clinical effect, even without data on statistical significance, was also advised.

Advice on assigning severity scores was also provided: despite the fact that some effects were detected by histochemical methods with no statistical data, a severity score was could be assigned when the histopathological lesion demonstrates a direct impact upon target organs. The outcome of fetal death was considered to be equally severe as death, with a severity score 9 and –8. In the case of limited reporting of outcomes by the authors (i.e. lack of quantitative information), behaviour changes with signs of aggressiveness were assigned a severity score of 2 instead of 6. Where local adverse effects were observed it was recognized that they depend on the chemical form of Mn, pH, and exposure pattern (e.g. nasal histopathology in inhalation toxicity studies); in such instances, these portal of entry (local) effects were assigned a score 5. In studies where health effects were scored following a recovery period, severity scores 8 and –7 were assigned when no recovery was observed.

2.4. Dose conversions

Reporting of Mn exposure levels is not uniform across studies pertaining to oral exposure. In some studies, Mn dose was expressed in mg Mn per kg body weight per day, while in others only concentrations in water or food were reported. Because a common dose metric is required for use in categorical regression of multiple studies, all Mn exposures were expressed in mg/kg bw/day. Mn concentrations in food or water were converted into Mn doses based on body weight and food/water consumption. The dose conversions were done as follows:

dose in mg/kg bw/day

$$= \frac{(\text{food intake in grams/day}) * [\text{Mn}] \text{in food in ppm or mg/kg diet}}{(\text{body weight in kg} * 1000)}$$

dose in mg/kg bw/day

$$= \frac{(\text{water in take in mL/day}) * [\text{Mn}] \text{in water in mg/mL} * 1000}{(\text{body weight in g})}$$

Concentrations of Mn in basal diet were converted to Mn doses using the same approach (US EPA, 2011). Many studies do not report Mn concentrations in basal diet; in such cases, Mn dose from the basal diet was assumed on the basis of existing data. The distribution of existing data on Mn doses from basal diet was examined visually for rats and mice, the two species with the greatest numbers of experiments in the database (Table 3). Due to

Table 1
The 18-point severity scoring matrix developed for application to the MnDB.

Direction of Effect	Severity Score	Description of Adverse Health Effect
Deficiency	-8	Death
	-7	Irreversible anatomic pathology
	-6	Clinical signs of deficiency, reversible anatomic pathology
	-5	Functional changes (e.g. alterations in reproductive, hepatic, renal or pancreatic function, changes in activity of pancreatic enzymes). Changes in bone density parameters
	-4	Metabolic perturbations. Changes in Fe, Cu, Zn tissue/biological fluids concentrations. Changes in bone metabolism (e.g. changes in activity of alkaline phosphatase) Changes in body or organ weight
	-3	Biochemical changes involved in pathways of manganese utilization reflecting the deficiency state (e.g. loss of Mn-dependent enzyme function). Decrease in tissue/biofluid Mn concentrations Changes comparable to those seen in category 3 excess
	-2	Changes of unknown clinical significance Changes in gene expression of Mn-dependent enzymes Changes comparable to those seen in category 2 excess
	-1	Decreased Mn excretion; increased gastrointestinal Mn absorption
	No Effect	0
Excess	1	Reduced gastrointestinal tract Mn absorption, increased Mn excretion, increase in liver and/or bile Mn concentrations
	2	Changes of unknown clinical significance Changes in gene or protein expression of transport proteins, antioxidant enzymes, neurotransmitter Changes in Mn concentrations in non-target organs/bio-fluids (e.g. kidney, blood, serum, urine) Changes in tissue Se and electrolyte concentrations (e.g. K, Mg, Na, Ca)
	3	Biochemical and/or cellular changes involved in manganese toxicity pathways Increased reactive oxygen species generation, decreased antioxidant enzyme activity Glial activation, increased levels of neuro-inflammatory markers Alteration in the level of neuro-transmitters Mitochondrial dysfunction, altered energy metabolism Increase in brain or lung (inhalation) Mn concentrations
	4	Metabolic perturbations Changes in Fe, Cu, Zn, tissue/biological fluids concentrations Decreased body weight; changes in organ weight Changes in responses to stimuli (e.g. amphetamine, cocaine, electroshock, immunological)
	5	Clinically significant functional changes (e.g. alterations in hepatic, renal, pulmonary, or reproductive function) Portal of entry (e.g. respiratory tract, gastrointestinal, dermal) anatomic pathology or related responses Neurological symptoms (e.g. mood changes, irritability)
	6	Adverse neurofunctional changes (electrophysiological, cognitive, and behavioral)
	7	Overt clinical signs of toxicity (e.g. tremors, seizures, ataxia)
	8	Irreversible anatomic pathology (e.g. neuronal death necrosis and apoptosis)
	9	Irreversible adverse neurological effects (e.g. "cock walk") Death

Bold values are severity scores used to characterize adverse health effects.

Table 2
Assumptions on Mn in Basal Diet.

Species	Mean Dose (mg/kg bw)	Median Dose (mg/kg bw)	Source(s)	Assumptions
Rat	4.5	4	Calculated from <i>MnDB</i>	N/A
Mouse	11	10	Calculated from <i>MnDB</i>	N/A
Monkey	3.3	N/A	Schroeter et al. (2012) US EPA (1988)	80 ppm concentration in basal diet 8 kg body weight 330 g/day food intake
Rabbit	3	N/A	http://www.sdsdiets.com/pdfs/rabbit-standard.pdf	90 ppm concentration in basal diet 3.8 kg body weight 120 g/day food intake
Guinea Pig	5	N/A	US EPA (1988)	80 ppm Mn in basal diet 500 g body weight 32 g/day food intake

presence of outliers, the median basal diet Mn concentration is preferable to the mean concentration.

In experiments where data on Mn in the basal diet were unavailable, median doses of Mn from basal diet were assigned according to the values provided below in Table 2.

3. Results

Each observation in the database corresponds to a single dose level from each study, with the severity score(s) corresponding to the adverse health outcome(s) seen at that dose. For each data point, information is provided on the species, sex, age, route of exposure, animal strain, exposure level, and duration of exposure. The database incorporates information from eight different

Table 3
Demographic Characteristics of the MnDB.

Characteristic		Number of studies/ Number of dose groups
Species	<i>Rattus norvegicus</i> (rat)	251/658
	<i>Mus musculus</i> (mouse)	73/197
	Monkey ^a	15/33
	<i>Homo sapiens</i> (human)	22/47
	<i>Sus scrofa domestica</i> (domesticated pig)	4/8
	<i>Oryctolagus cuniculus</i> (domesticated rabbit)	10/26
	<i>Mesocricetus auratus</i> (Syrian hamster)	1/2
	<i>Cavia porcellus</i> (domesticated guinea pig)	2/4
Sex	Male	254/640
	Female	72/201
	Both sexes	30/85
	Unknown	22/48
Exposure route	Oral	323/827
	Drinking water	87/198
	Food	138/369
	Gavage	94/253
	Tablet or capsule	4/8
	Inhalation	55/147
Type of study	Experimental	367/954
	Observational	11/20

^a Rhesus (*Macaca mulatta*), cynomolgus (*Macaca fascicularis*), and squirrel monkeys (*Saimiri sciureus*) were coded as one species (monkey) in the database.

species, males and females of all ages, inhalation and oral exposure routes, as well as experimental and observational studies, providing a comprehensive repository of information for exposure-response assessment.

3.1. Distribution of study characteristics in the MnDB

Table 3 presents the characteristics of these studies by species, sex, exposure route, and study type. The data summarized in this

table provides the raw data needed for categorical regression analysis.

Table 3 demonstrates the vast majority of studies included in the database were performed on rodents, with males, and via the oral route of exposure. Studies on humans tend to focus on marginal to moderate effects due to both Mn deficiency and excess. In contrast, animal studies tend to focus primarily on more severe effects, with the objective of defining a broad continuum of toxicity. At this time, human data are limited, and may be inadequate for the application of categorical regression, with convergence issues due to complete separation or quasi-separation (Allison, 2004) likely to be encountered as artifacts of a small data set. Complete separation occurs when there is one exposure level, C , that perfectly separates the data. In this case, one can ascertain that for exposure levels less than C , $Y=0$, and for exposure levels greater than C , $Y=1$. Quasi-separation occurs when exposure level C yields $Y=0$ and $Y=1$; this often occurs when exposure-response data from different studies are combined. As a consequence, human and animal data will likely need to be combined when conducting categorical regression analysis. The *CatReg* software permits model parameters to be stratified by animal species: a categorical regression model can be parameterized so that human data are used to estimate the intercept, while animal data are used to characterize the slope (Haber et al., 2001).

3.2. Distribution of severity scores for oral exposure data

The common response scale was applied to the MnDB. In the data abstraction stage, the group size from each experiment was also recorded. In determining the distribution of severity scores, the group level entries were converted to individual level entries, and their distributions are presented above in Table 4, which highlights the extensive data available for excess exposures. By comparison, the information available for deficiency exposure is much more limited. Within the database, it is clear the number of studies on excess exposures to Mn is far greater than the number of studies on deficiency. This reflects the information currently available in the scientific literature, and corresponds to the greater regulatory concern about Mn excess than Mn deficiency.

Table 4
Distribution of severity scores based on oral exposure data in the MnDB.

Excess or Deficiency	Severity Score	Species								
		Humans	Monkeys	Rats	Mice	Hamsters	Guinea Pigs	Rabbits	Pigs	Total
Deficiency	-8	0	0	0	0	0	0	0	0	0
	-7	0	0	22	0	0	0	34	26	82
	-6	0	0	8	12	0	0	0	0	20
	-5	0	0	36	0	0	0	0	6	42
	-4	0	0	163	14	0	0	0	0	177
	-3	0	0	217	0	0	0	0	0	217
	-2	0	0	36	0	0	0	0	0	36
	-1	4	0	16	0	0	0	0	0	20
No effect	0	71	16	3382	1112	16	31	116	32	4776
Excess	1	0	0	18	0	0	0	0	0	18
	2	34	0	353	173	0	12	32	0	604
	3	26	0	555	86	0	0	0	16	683
	4	0	4	618	230	0	0	0	0	852
	5	0	0	53	14	0	0	0	0	67
	6	0	16	470	75	0	0	0	0	561
	7	0	0	39	354	0	0	0	0	393
	8	0	4	31	34	0	0	0	0	69
	9	0	0	96	0	0	0	0	0	96
Total		135	36	6113	2104	16	43	182	80	8713

4. Discussion

An important contribution of this work was the development of an 18-point severity scoring matrix designed to standardize health endpoints onto a common scale for the application of categorical regression. This matrix can be adopted as a general template for all metals. Since all metals exhibit different toxicological properties, this general template could be modified to accommodate the characteristics of the metal under study, providing a stepping stone to begin to look at essential elements known to exhibit both health benefits and health risks that may be balanced using categorical regression modeling techniques.

The *MnDB* offers the largest, most current library of data abstracted from relevant Mn studies for exposure-response assessment. The database has proven effective as an organizational tool to synthesize information abstracted from scientific articles. A review of the database reveals considerable diversity among the available studies with regards to species, route of exposure, sex, and age, indicating stratification is an essential aspect in the categorical regression analysis. The database is also useful for identifying gaps in the literature, such as the limited amount of data on Mn toxicity due to deficiency. Future directions for this work include more accurate exposure characterization; one such example is developing biomarkers which can be used to quantitate exposure. As additional information on Mn toxicity due to deficiency accrues in the future, a more complete description of the U-shaped dose response curve for Mn as an essential element demonstrating toxicity due to both excess and deficiency may be possible.

Conflict of interest

None.

Transparency document

The [Transparency document](#) associated with this article can be found in the online version.

Appendix A. List of Eligible Articles

[1] Abdel Rahim AG, Arthur JR, Mills CF. Effects of dietary copper, cadmium, iron, molybdenum and manganese on selenium utilization by the rat. *J Nutr* 1986; 116(3): 403–411.

[2] Adkins B, Jr., Luginbuhl GH, Gardner DE. Biochemical changes in pulmonary cells following manganese oxide inhalation. *J Toxicol Environ Health* 1980; 6(2): 445–454.

[3] Adkins B, Jr., Luginbuhl GH, Miller FJ, et al. Increased pulmonary susceptibility to streptococcal infection following inhalation of manganese oxide. *Environ Res* 1980; 23(1): 110–120.

[4] Adkins B, Jr., Luginbuhl GH, Gardner DE. Acute exposure of laboratory mice to manganese oxide. *Am Ind Hyg Assoc J* 1980; 41(7): 494–500.

[5] Albiin N, Kartalis N, Bergquist A, et al. Manganese chloride tetrahydrate (CMC-001) enhanced liver MRI: evaluation of efficacy and safety in healthy volunteers. *MAGMA* 2012; 25(5): 361–368.

[6] Ali MM, Murthy RC, Saxena DK, et al. Effect of low protein diet on manganese neurotoxicity: II. Brain GABA and seizure susceptibility. *Neurobehav Toxicol Teratol* 1983; 5(3): 385–389.

[7] Ali MM, Murthy RC, Mandal SK, et al. Effect of low protein diet on manganese neurotoxicity: III. Brain neurotransmitter levels. *Neurobehav Toxicol Teratol* 1985; 7(5): 427–431.

[8] Anderson JG, Cooney PT, Erikson KM. Brain manganese accumulation is inversely related to gamma-amino butyric acid uptake in male and female rats. *Toxicol Sci* 2007; 95(1): 188–195.

[9] Anderson JG, Cooney PT, Erikson KM. Inhibition of DAT function attenuates manganese accumulation in the globus pallidus. *Environ Toxicol Pharmacol* 2007; 23(2): 179–184.

[10] Anderson JG, Fordahl SC, Cooney PT, et al. Manganese exposure alters extracellular GABA, GABA receptor and transporter protein and mRNA levels in the developing rat brain. *Neurotoxicology* 2008; 29(6): 1044–1053.

[11] Anderson JG, Fordahl SC, Cooney PT, et al. Extracellular norepinephrine, norepinephrine receptor and transporter protein and mRNA levels are differentially altered in the developing rat brain due to dietary iron deficiency and manganese exposure. *Brain Res* 2009; 1281: 1–14.

[12] Antonini JM, Sriram K, Benkovic SA, et al. Mild steel welding fume causes manganese accumulation and subtle neuro-inflammatory changes but not overt neuronal damage in discrete brain regions of rats after short-term inhalation exposure. *Neurotoxicology* 2009; 30(6): 915–925.

[13] Antunes MB, Bowler R, Doty RL. San Francisco/Oakland Bay Bridge Welder Study: olfactory function. *Neurology* 2007; 69(12): 1278–1284.

[14] Apgar J. Comparison of the effect of copper, manganese, and zinc deficiencies on parturition in the rat. *Am J Physiol* 1968; 215(6): 1478–1481.

[15] Avila DS, Gubert P, Fachinetti R, et al. Involvement of striatal lipid peroxidation and inhibition of calcium influx into brain slices in neurobehavioral alterations in a rat model of short-term oral exposure to manganese. *Neurotoxicology* 2008; 29(6): 1062–1068.

[16] Avila DS, Colle D, Gubert P, et al. A possible neuroprotective action of a vinyllic telluride against Mn-induced neurotoxicity. *Toxicol Sci* 2010; 115(1): 194–201.

[17] Bae YJ, Kim MH. Manganese supplementation improves mineral density of the spine and femur and serum osteocalcin in rats. *Biol Trace Elem Res* 2008; 124(1): 28–34.

[18] Baly DL, Curry DL, Keen CL, et al. Effect of manganese deficiency on insulin secretion and carbohydrate homeostasis in rats. *J Nutr* 1984; 114(8): 1438–1446.

[19] Bast-Petersen R, Ellingsen DG, Hetland SM, et al. Neuropsychological function in manganese alloy plant workers. *Int Arch Occup Environ Health* 2004; 77(4): 277–287.

[20] Bataineh HN, Bataineh ZM, Daradka H. Short-term exposure of female rats to industrial metal salts: Effect on implantation and pregnancy. *Reproductive Medicine and Biology* 2007; 6(3): 179–183.

[21] Bergstrom R. Acute pulmonary toxicity of manganese dioxide. *Scand J Work Environ Health* 1977; 3 Suppl. 1: 1–41.

[22] Bird ED, Anton AH, Bullock B. The effect of manganese inhalation on basal ganglia dopamine concentrations in rhesus monkey. *Neurotoxicology* 1984; 5(1): 59–65.

[23] Bonilla E, Diez-Ewald M. Role of submaxillary glands in radiomanganese metabolism. *Experientia* 1972; 28(10): 1152–1153.

[24] Bonilla E, Diez-Ewald M. Effect of L-DOPA on brain concentration of dopamine and homovanillic acid in rats after chronic manganese chloride administration. *J Neurochem* 1974; 22(2): 297–299.

[25] Bonilla E. Increased GABA content in caudate nucleus of rats after chronic manganese chloride administration. *J Neurochem* 1978; 31(2): 551–552.

[26] Bonilla E. L-tyrosine hydroxylase activity in the rat brain after chronic oral administration of manganese chloride. *Neurobehav Toxicol* 1980; 2(1): 37–41.

[27] Bonilla E. Chronic manganese intake induces changes in the motor activity of rats. *Exp Neurol* 1984; 84(3): 696–700.

- [28] Bonilla E, Prasad AL. Effects of chronic manganese intake on the levels of biogenic amines in rat brain regions. *Neurobehav Toxicol Teratol* 1984; 6(5): 341–344.
- [29] Bonilla E, Prasad ALN, Davila JO, et al. Free amino acids in plasma and brain after chronic manganese intake in rats. *Investigacion Clinica* 1988; 29(2): 55–60.
- [30] Bouchard M, Mergler D, Baldwin M, et al. Neuropsychiatric symptoms and past manganese exposure in a ferro-alloy plant. *Neurotoxicology* 2007; 28(2): 290–297.
- [31] Bouchard M, Mergler D, Baldwin M, et al. Neurobehavioral functioning after cessation of manganese exposure: a follow-up after 14 years. *Am J Ind Med* 2007; 50(11): 831–840.
- [32] Bouchard M, Mergler D, Baldwin ME, et al. Manganese cumulative exposure and symptoms: a follow-up study of alloy workers. *Neurotoxicology* 2008; 29(4): 577–583.
- [33] Bowler RM, Nakagawa S, Drezgic M, et al. Sequelae of fume exposure in confined space welding: a neurological and neuropsychological case series. *Neurotoxicology* 2007; 28(2): 298–311.
- [34] Bowler RM, Roels HA, Nakagawa S, et al. Dose-effect relationships between manganese exposure and neurological, neuropsychological and pulmonary function in confined space bridge welders. *Occup Environ Med* 2007; 64(3): 167–177.
- [35] Bowler RM, Nakagawa S, Drezgic M, et al. Sequelae of fume exposure in confined space welding: a neurological and neuropsychological case series. *Neurotoxicology* 2007; 28(2): 298–311.
- [36] Bowler RM, Gocheva V, Harris M, et al. Prospective study on neurotoxic effects in manganese-exposed bridge construction welders. *Neurotoxicology* 2011; 32(5): 596–605.
- [37] Brannon PM, Collins VP, Korc M. Alterations of pancreatic digestive enzyme content in the manganese-deficient rat. *J Nutr* 1987; 117(2): 305–311.
- [38] Bredow S, Falgout MM, March TH, et al. Subchronic inhalation of soluble manganese induces expression of hypoxia-associated angiogenic genes in adult mouse lungs. *Toxicol Appl Pharmacol* 2007; 221(2): 148–157.
- [39] Brenneman KA, Cattley RC, Ali SF, et al. Manganese-induced developmental neurotoxicity in the CD rat: is oxidative damage a mechanism of action? *Neurotoxicology* 1999; 20(2–3): 477–487.
- [40] Brock AA, Chapman SA, Ulman EA, et al. Dietary manganese deficiency decreases rat hepatic arginase activity. *J Nutr* 1994; 124(3): 340–344.
- [41] Buchet JP, Magos C, Roels H, et al. Urinary excretion of homovanillic acid in workers exposed to manganese. *Int Arch Occup Environ Health* 1993; 65(2): 131–133.
- [42] Burch RE, Williams RV, Hahn HK, et al. Tissue trace element and enzyme content in pigs fed a low manganese diet. I. A relationship between manganese and selenium. *J Lab Clin Med* 1975; 86(1): 132–139.
- [43] Calabresi P, Ammassari-Teule M, Gubellini P, et al. A synaptic mechanism underlying the behavioral abnormalities induced by manganese intoxication. *Neurobiol Dis* 2001; 8(3): 419–412.
- [44] Camner P, Curstedt T, Jarstrand C, et al. Rabbit lung after inhalation of manganese chloride: a comparison with the effects of chlorides of nickel, cadmium, cobalt, and copper. *Environ Res* 1985; 38(2): 301–309.
- [45] Cavallari JM, Eisen EA, Fang SC, et al. PM2.5 metal exposures and nocturnal heart rate variability: a panel study of boilermaker construction workers. *Environ Health* 2008; 7: 36.
- [46] Centonze D, Gubellini P, Bernardi G, et al. Impaired excitatory transmission in the striatum of rats chronically intoxicated with manganese. *Exp Neurol* 2001; 172(2): 469–476.
- [47] Chandra SV, Tandon SK. Enhanced manganese toxicity in iron-deficient rats. *Environ Physiol Biochem* 1973; 3: 230–235.
- [48] Chandra SV, Imam Z. Manganese induced histochemical and histological alterations in gastrointestinal mucosa of guinea pigs. *Acta Pharmacol Toxicol (Copenh)* 1973; 33(5): 449–458.
- [49] Chandra SV, Shukla GS. Manganese encephalopathy in growing rats. *Environ Res* 1978; 15(1): 28–37.
- [50] Chandra SV, Srivastava RS, Shukla GS. Regional distribution of metals and biogenic amines in the brain of monkeys exposed to manganese. *Toxicology Letters* 1979; 4: 189–192.
- [51] Chandra SV, Shukla GS. Concentrations of striatal catecholamines in rats given manganese chloride through drinking water. *J Neurochem* 1981; 36(2): 683–687.
- [52] Chang SC, Brannon PM, Korc M. Effects of dietary manganese deficiency on rat pancreatic amylase mRNA levels. *J Nutr* 1990; 120(10): 1228–1234.
- [53] Chtourou Y, Fetoui H, Sefi M, et al. Silymarin, a natural antioxidant, protects cerebral cortex against manganese-induced neurotoxicity in adult rats. *Biometals* 2010; 23(6): 985–996.
- [54] Chtourou Y, Fetoui H, Garoui eM, et al. Improvement of cerebellum redox States and cholinergic functions contribute to the beneficial effects of silymarin against manganese-induced neurotoxicity. *Neurochem Res* 2012; 37(3): 469–479.
- [55] Clegg MS, Donovan SM, Monaco MH, et al. The influence of manganese deficiency on serum IGF-1 and IGF binding proteins in the male rat. *Proc Soc Exp Biol Med* 1998; 219(1): 41–47.
- [56] Colin-Barenque L, Souza-Gallardo LM, Fortoul TI. Toxic effects of inhaled manganese on the olfactory bulb: an ultrastructural approach in mice. *J Electron Microscop (Tokyo)* 2011; 60(1): 73–78.
- [57] Coulston F, Griffin T. Inhalation toxicology of airborne particulate manganese in Rhesus monkeys. EPA-600/1-77-026. 1977. US EPA. <http://nepis.epa.gov/Exe/ZyNET.exe/91013HKS.TXT?ZyActionD=ZyDocument&Client=EPA&Index=1976±-Thru±1980&Docs=&Query=&Time=&EndTime=&SearchMethod=1&TocRestrict=n&Toc=&TocEntry=&QField=&QFieldYear=&QFieldMonth=&QFieldDay=&IntQFieldOp=0&ExtQFieldOp=0&XmlQuery=&File=D%3A%5Czyfiles%5CIndex%20Data%5C76thru80%5CTxt%5C00000022%5C91013HKS.txt&User=ANONYMOUS&Password=anonymous&SortMethod=h%7C-&MaximumDocuments=1&FuzzyDegree=0&ImageQuality=r75g8/r75g8/x150y150g16/i425&Display=p%7C&DefSeekPage=x&SearchBack=ZyActionL&Back=ZyActionS&BackDesc=Results%20page&MaximumPages=1&ZyEntry=1&SeekPage=x&ZyPURL#>.
- [58] Davis CD, Ney DM, Greger JL. Manganese, iron and lipid interactions in rats. *J Nutr* 1990; 120(5): 507–513.
- [59] Davis CD, Greger JL. Longitudinal changes of manganese-dependent superoxide dismutase and other indexes of manganese and iron status in women. *Am J Clin Nutr* 1992; 55(3): 747–752.
- [60] Deskin R, Bursian SJ, Edens FW. The effect of chronic manganese administration on some neurochemical and physiological variables in neonatal rats. *Gen Pharmacol* 1981; 12(4): 279–280.
- [61] Deskin R, Bursian SJ, Edens FW. Neurochemical alterations induced by manganese chloride in neonatal rats. *Neurotoxicology* 1981; 2(1): 65–73.
- [62] Desole MS, Miele M, Esposito G, et al. Monoaminergic systems activity and cellular defense mechanisms in the brainstem of young and aged rats subchronically exposed to manganese. *Neurosci Lett* 1994; 177(1–2): 71–74.
- [63] Desole MS, Miele M, Esposito G, et al. Dopaminergic system activity and cellular defense mechanisms in the striatum and striatal synaptosomes of the rat subchronically exposed to manganese. *Arch Toxicol* 1994; 68(9): 566–570.
- [64] Desole MS, Esposito G, Migheli R, et al. Cellular defense mechanisms in the striatum of young and aged rats subchronically exposed to manganese. *Neuropharmacology* 1995; 34(3): 289–295.

- [65] Desole MS, Esposito G, Migheli R, et al. Glutathione deficiency potentiates manganese toxicity in rat striatum and brainstem and in PC12 cells. *Pharmacol Res* 1997; 36(4): 285–292.
- [66] Desole MS, Serra PA, Esposito G, et al. Glutathione deficiency potentiates manganese-induced increases in compounds associated with high-energy phosphate degradation in discrete brain areas of young and aged rats. *Aging (Milano)* 2000; 12(6): 470–477.
- [67] Dobson AW, Weber S, Dorman DC, et al. Oxidative stress is induced in the rat brain following repeated inhalation exposure to manganese sulfate. *Biol Trace Elem Res* 2003; 93(1–3): 113–126.
- [68] Dorman DC, Struve MF, Vitarella D, et al. Neurotoxicity of manganese chloride in neonatal and adult CD rats following subchronic (21-day) high-dose oral exposure. *J Appl Toxicol* 2000; 20(3): 179–187.
- [69] Dorman DC, Struve MF, James RA, et al. Influence of dietary manganese on the pharmacokinetics of inhaled manganese sulfate in male CD rats. *Toxicol Sci* 2001; 60(2): 242–251.
- [70] Dorman DC, Struve MF, James RA, et al. Influence of dietary manganese on the pharmacokinetics of inhaled manganese sulfate in male CD rats. *Toxicol Sci* 2001; 60(2): 242–251.
- [71] Dorman DC, Struve MF, Wong BA. Brain manganese concentrations in rats following manganese tetroxide inhalation are unaffected by dietary manganese intake. *Neurotoxicology* 2002; 23(2): 185–195.
- [72] Dorman DC, McManus BE, Parkinson CU, et al. Nasal toxicity of manganese sulfate and manganese phosphate in young male rats following subchronic (13-week) inhalation exposure. *Inhal Toxicol* 2004; 16(6–7): 481–488.
- [73] Dorman DC, Struve MF, Gross EA, et al. Sub-chronic inhalation of high concentrations of manganese sulfate induces lower airway pathology in rhesus monkeys. *Respir Res* 2005; 6: 121.
- [74] Dorman DC, Struve MF, Marshall MW, et al. Tissue manganese concentrations in young male rhesus monkeys following subchronic manganese sulfate inhalation. *Toxicol Sci* 2006; 92(1): 201–210.
- [75] Dorman DC, Struve MF, Wong BA, et al. Correlation of brain magnetic resonance imaging changes with pallidum manganese concentrations in rhesus monkeys following subchronic manganese inhalation. *Toxicol Sci* 2006; 92(1): 219–227.
- [76] Dorman DC, Struve MF, Norris A, et al. Metabolomic analyses of body fluids after subchronic manganese inhalation in rhesus monkeys. *Toxicol Sci* 2008; 106(1): 46–54.
- [77] Egbe-Nwiyi TN, Aliyu MM, Igbokwe IO. Effects of oral supplementation with manganese chloride on the severity of *Trypanosoma brucei* and *Trypanosoma congolense* infections in rats. *African Journal of Biomedical Research* 2010; 13(1): 27–31.
- [78] Elbetieha A, Bataineh H, Darmani H, et al. Effects of long-term exposure to manganese chloride on fertility of male and female mice. *Toxicol Lett* 2001; 119(3): 193–201.
- [79] Ellingsen DG, Haug E, Gaarder PI, et al. Endocrine and immunologic markers in manganese alloy production workers. *Scand J Work Environ Health* 2003; 29(3): 230–238.
- [80] Ellingsen DG, Haug E, Ulvik RJ, et al. Iron status in manganese alloy production workers. *J Appl Toxicol* 2003; 23(4): 239–247.
- [81] Ellingsen DG, Konstantinov R, Bast-Pettersen R, et al. A neurobehavioral study of current and former welders exposed to manganese. *Neurotoxicology* 2008; 29(1): 48–59.
- [82] Ellis GH, Smith SE, Gates EM. Further studies of manganese deficiency in the rabbit. *J Nutr* 1947; 34(1): 21–31.
- [83] Erikson KM, Dorman DC, Lash LH, et al. Airborne manganese exposure differentially affects end points of oxidative stress in an age- and sex-dependent manner. *Biol Trace Elem Res* 2004; 100(1): 49–62.
- [84] Erikson KM, Dorman DC, Lash LH, et al. Manganese inhalation by rhesus monkeys is associated with brain regional changes in biomarkers of neurotoxicity. *Toxicol Sci* 2007; 97(2): 459–466.
- [85] Erikson KM, Dorman DC, Lash LH, et al. Duration of airborne-manganese exposure in rhesus monkeys is associated with brain regional changes in biomarkers of neurotoxicity. *Neurotoxicology* 2008; 29(3): 377–385.
- [86] Eriksson H, Lenngren S, Heilbronn E. Effect of long-term administration of manganese on biogenic amine levels in discrete striatal regions of rat brain. *Arch Toxicol* 1987; 59(6): 426–431.
- [87] Everson GJ, Daniels AL. A study of manganese retentions in children. *J Nutr* 1934; 8(5): 497–502.
- [88] Exon JH, Koller LD. Effects of feeding manganese antiknock gasoline additive exhaust residues (Mn₃O₄) in rats. *Bull Environ Contam Toxicol* 1975; 14(3): 370–373.
- [89] Fahim FA, Morcos NY, Esmat AY. Effects of dietary magnesium and/or manganese variables on the growth rate and metabolism of mice. *Ann Nutr Metab* 1990; 34(3): 183–192.
- [90] Finley JW, Davis CD. Manganese absorption and retention in rats is affected by the type of dietary fat. *Biol Trace Elem Res* 2001; 82(1–3): 143–158.
- [91] Finley JW, Penland JG, Pettit RE, et al. Dietary manganese intake and type of lipid do not affect clinical or neuropsychological measures in healthy young women. *J Nutr* 2003; 133(9): 2849–2856.
- [92] Fordahl S, Cooney P, Qiu Y, et al. Waterborne manganese exposure alters plasma, brain, and liver metabolites accompanied by changes in stereotypic behaviors. *Neurotoxicol Teratol* 2012; 34(1): 27–36.
- [93] Fordahl SC, Anderson JG, Cooney PT, et al. Manganese exposure inhibits the clearance of extracellular GABA and influences taurine homeostasis in the striatum of developing rats. *Neurotoxicology* 2010; 31(6): 639–646.
- [94] Freeland-Graves JH, Behmardi F, Bales CW, et al. Metabolic balance of manganese in young men consuming diets containing five levels of dietary manganese. *J Nutr* 1988; 118(6): 764–773.
- [95] Freundt KJ, Ibrahim HA. Growth of rats during a subchronic intake of the heavy metals Pb, Cd, Zn, Mn, Cu, Hg, and Be. *Pol J Occup Med* 1990; 3(2): 227–232.
- [96] Friedman BJ, Freeland-Graves JH, Bales CW, et al. Manganese balance and clinical observations in young men fed a manganese-deficient diet. *J Nutr* 1987; 117(1): 133–143.
- [97] Gaillard E, Laurant P, Robin S, et al. Effects of long-term high manganese intake on magnesium metabolism in rats. *Magn Res* 1996; 9(2): 119–123.
- [98] Gianutsos G, Murray MT. Alterations in brain dopamine and GABA following inorganic or organic manganese administration. *Neurotoxicology* 1982; 3(3): 75–81.
- [99] Golub MS, Han B, Keen CL, et al. Effects of dietary aluminum excess and manganese deficiency on neurobehavioral endpoints in adult mice. *Toxicol Appl Pharmacol* 1992; 112(1): 154–160.
- [100] Golub MS, Hogrefe CE, Germann SL, et al. Neurobehavioral evaluation of rhesus monkey infants fed cow's milk formula, soy formula, or soy formula with added manganese. *Neurotoxicol Teratol* 2005; 27(4): 615–627.
- [101] Gong H, Amemiya T. Ultrastructure of retina of manganese-deficient rats. *Invest Ophthalmol Vis Sci* 1996; 37(10): 1967–1974.
- [102] Gupta A, Shukla GS. Enzymatic antioxidants in erythrocytes following heavy metal exposure: possible role in early diagnosis of poisoning. *Bull Environ Contam Toxicol* 1997; 58(2): 198–205.
- [103] Gupta SK, Murthy RC, Chandra SV. Neuromelanin in manganese-exposed primates. *Toxicol Lett* 1980; 6(1): 17–20.

- [104] HaMai D, Rinderknecht AL, Guo-Sharman K, et al. Decreased expression of inflammation-related genes following inhalation exposure to manganese. *Neurotoxicology* 2006; 27 (3): 395–401.
- [105] Hastings CE, Jr., Llewellyn GC. Reduced aflatoxicosis in livers of hamsters fed a manganese sulfate supplement. *Nutr Cancer* 1987; 10(1–2): 67–77.
- [106] Hietanen E, Kilpio J, Savolainen H. Neurochemical and biotransformational enzyme responses to manganese exposure in rats. *Arch Environ Contam Toxicol* 1981; 10 (3): 339–345.
- [107] Hiney JK, Srivastava VK, Dees WL. Manganese induces IGF-1 and cyclooxygenase-2 gene expressions in the basal hypothalamus during prepubertal female development. *Toxicol Sci* 2011; 121 (2): 389–396.
- [108] Horvath E, Mate Z, Takacs S, et al. General and Electrophysiological Toxic Effects of Manganese in Rats following Subacute Administration in Dissolved and Nanoparticle Form. *ScientificWorldJournal* 2012; 2012: 520632.
- [109] Jarvinen R, Ahlstrom A. Effect of the dietary manganese level on tissue manganese, iron, copper and zinc concentrations in female rats and their fetuses. *Med Biol* 1975; 53(2): 93–99.
- [110] Kalea AZ, Harris PD, Klimis-Zacas DJ. Dietary manganese suppresses alpha1 adrenergic receptor-mediated vascular contraction. *J Nutr Biochem* 2005; 16(1): 44–49.
- [111] Kalea AZ, Lamari FN, Theocharis AD, et al. Dietary manganese affects the concentration, composition and sulfation pattern of heparan sulfate glycosaminoglycans in Sprague-Dawley rat aorta. *Biometals* 2006; 19(5): 535–546.
- [112] Kawada J, Nishida M, Yoshimura Y, et al. Manganese ion as a goitrogen in the female mouse. *Endocrinol Jpn* 1985; 32(5): 635–643.
- [113] Kawano J, Ney DM, Keen CL, et al. Altered high density lipoprotein composition in manganese-deficient Sprague-Dawley and Wistar rats. *J Nutr* 1987; 117 (5): 902–906.
- [114] Kern CH, Stanwood GD, Smith DR. Prewaning manganese exposure causes hyperactivity, disinhibition, and spatial learning and memory deficits associated with altered dopamine receptor and transporter levels. *Synapse* 2010; 64(5): 363–378.
- [115] Kern CH, Smith DR. Prewaning Mn exposure leads to prolonged astrocyte activation and lasting effects on the dopaminergic system in adult male rats. *Synapse* 2011; 65(6): 532–544.
- [116] Khandelwal S, Tandon SK. Effect of manganese on certain enzymes and constituents of blood and serum in rabbits. II. *Environ Res* 1981; 24(1): 82–88.
- [117] Kim HY, Lee CK, Lee JT, et al. Effects of manganese exposure on dopamine and prolactin production in rat. *Neuroreport* 2009; 20(1): 69–73.
- [118] Kimura M, Yagi N, Itokawa Y. Effect of subacute manganese feeding on serotonin metabolism in the rat. *J Toxicol Environ Health* 1978; 4(5–6): 701–707.
- [119] Klimis-Tavantzis DJ, Leach RM, Jr., Kris-Etherton PM. The effect of dietary manganese deficiency on cholesterol and lipid metabolism in the Wistar rat and in the genetically hypercholesterolemic RICO rat. *J Nutr* 1983; 113(2): 328–336.
- [120] Klimis-Tavantzis DJ, Taylor PN, Lewis RA, et al. Effects of dietary manganese deficiency on high density lipoprotein composition and metabolism in Sprague-Dawley rats. *Nutrition Research* 1993; 13(8): 953–968.
- [121] Komura J, Sakamoto M. Short-term oral administration of several manganese compounds in mice: physiological and behavioral alterations caused by different forms of manganese. *Bull Environ Contam Toxicol* 1991; 46(6): 921–928.
- [122] Komura J, Sakamoto M. Effects of manganese forms on biogenic amines in the brain and behavioral alterations in the mouse: long-term oral administration of several manganese compounds. *Environ Res* 1992; 57(1): 34–44.
- [123] Komura J, Sakamoto M. Chronic oral administration of methylcyclopentadienyl manganese tricarbonyl altered brain biogenic amines in the mouse: comparison with inorganic manganese. *Toxicol Lett* 1994; 73(1): 65–73.
- [124] Kontur PJ, Fechter LD. Brain regional manganese levels and monoamine metabolism in manganese-treated neonatal rats. *Neurotoxicol Teratol* 1988; 10(4): 295–303.
- [125] Kristensson K, Eriksson H, Lundh B, et al. Effects of manganese chloride on the rat developing nervous system. *Acta Pharmacol Toxicol (Copenh)* 1986; 59(5): 345–348.
- [126] Kumar M, Kannan A, Upreti RK, et al. Toxic Interaction of Lathyrus sativus and Manganese in Guinea Pig Intestine. *Toxicol Mech Methods* 2003; 13(4): 295–300.
- [127] Laohaudomchok W, Lin X, Herrick RF, et al. Neuropsychological effects of low-level manganese exposure in welders. *Neurotoxicology* 2011; 32(2): 171–179.
- [128] Laskey JW, Rehnberg GL, Hein JF, et al. Assessment of the male reproductive system in the preweaning rat following Mn3O4 exposure. *J Toxicol Environ Health* 1985; 15(2): 339–350.
- [129] Laurant P, Chanut E, Bobillier-Chaumont S, et al. Attenuation of the development of DOCA salt hypertension by a high Mn intake in the rat. *Trace Elements and Electrolytes* 2003; 20 (3): 172–180.
- [130] Lee B, Pine M, Johnson L, et al. Manganese acts centrally to activate reproductive hormone secretion and pubertal development in male rats. *Reprod Toxicol* 2006; 22(4): 580–585.
- [131] Lee DY, Johnson PE. Factors affecting absorption and excretion of ⁵⁴Mn in rats. *J Nutr* 1988; 118(12): 1509–1516.
- [132] Lee DY, Johnson PE. ⁵⁴Mn absorption and excretion in rats fed soy protein and casein diets. *Proc Soc Exp Biol Med* 1989; 190 (2): 211–216.
- [133] Lewis J, Bench G, Myers O, et al. Trigeminal uptake and clearance of inhaled manganese chloride in rats and mice. *Neurotoxicology* 2005; 26(1): 113–123.
- [134] Li GJ, Choi BS, Wang X, et al. Molecular mechanism of distorted iron regulation in the blood-CSF barrier and regional blood-brain barrier following in vivo subchronic manganese exposure. *Neurotoxicology* 2006; 27(5): 737–744.
- [135] Lipe GW, Duhart H, Newport GD, et al. Effect of manganese on the concentration of amino acids in different regions of the rat brain. *J Environ Sci Health B* 1999; 34(1): 119–132.
- [136] Liu X, Sullivan KA, Madl JE, et al. Manganese-induced neurotoxicity: the role of astroglial-derived nitric oxide in striatal interneuron degeneration. *Toxicol Sci* 2006; 91(2): 521–531.
- [137] Lucchini R, Bergamaschi E, Smargiassi A, et al. Motor function, olfactory threshold, and hematological indices in manganese-exposed ferroalloy workers. *Environ Res* 1997; 73(1–2): 175–180.
- [138] Lucchini R, Apostoli P, Perrone C, et al. Long-term exposure to “low levels” of manganese oxides and neurofunctional changes in ferroalloy workers. *Neurotoxicology* 1999; 20(2–3): 287–297.
- [139] Magour S, Maser H, Steffen I. Effect of daily oral intake of manganese on free polysomal protein synthesis of rat brain. *Acta Pharmacol Toxicol (Copenh)* 1983; 53(2): 88–91.
- [140] Maigetter RZ, Ehrlich R, Fenters JD, et al. Potentiating effects of manganese dioxide on experimental respiratory infections. *Environ Res* 1976; 11(3): 386–391.
- [141] Malecki EA, Huttner DL, Greger JL. Manganese status, gut endogenous losses of manganese, and antioxidant enzyme activity in rats fed varying levels of manganese and fat. *Biol Trace Elem Res* 1994; 42(1): 17–29.
- [142] Malecki EA, Greger JL. Manganese protects against heart mitochondrial lipid peroxidation in rats fed high levels of polyunsaturated fatty acids. *J Nutr* 1996; 126(1): 27–33.

- [143] Malecki EA, Radzanowski GM, Radzanowski TJ, et al. Biliary manganese excretion in conscious rats is affected by acute and chronic manganese intake but not by dietary fat. *J Nutr* 1996; 126(2): 489–498.
- [144] Martinez H, Bonilla E. Water intake and brain choline-acetyltransferase and acetylcholinesterase activities in manganese treated rats. *Neurobehav Toxicol Teratol* 1981; 3(3): 277–280.
- [145] Martins EN, Pessano NT, Leal L, et al. Protective effect of *Melissa officinalis* aqueous extract against Mn-induced oxidative stress in chronically exposed mice. *Brain Res Bull* 2012; 87(1): 74–79.
- [146] McCoy JH, Kenney MA, Gillham B. Immune response in rats fed marginal, adequate and high intakes of manganese. *Nutrition Reports International* 1979; 19(2): 165–172.
- [147] McDougall SA, Reichel CM, Farley CM, et al. Postnatal manganese exposure alters dopamine transporter function in adult rats: Potential impact on nonassociative and associative processes. *Neuroscience* 2008; 154(2): 848–860.
- [148] McDougall SA, Der-Ghazarian T, Britt CE, et al. Postnatal manganese exposure alters the expression of D2L and D2S receptor isoforms: relationship to PKA activity and Akt levels. *Synapse* 2011; 65(7): 583–591.
- [149] Mergler D, Huel G, Bowler R, et al. Nervous system dysfunction among workers with long-term exposure to manganese. *Environ Res* 1994; 64(2): 151–180.
- [150] Miele M, Serra PA, Esposito G, et al. Glutamate and catabolites of high-energy phosphates in the striatum and brainstem of young and aged rats subchronically exposed to manganese. *Aging (Milano)* 2000; 12(5): 393–397.
- [151] Miller KB, Caton JS, Finley JW. Manganese depresses rat heart muscle respiration. *Biofactors* 2006; 28(1): 33–46.
- [152] Missy P, Lanhers M-C, Cunat L, et al. Effects of subchronic exposure to manganese chloride on tissue distribution of three essential elements in rats. *International Journal of Toxicology* 2000; 19(5): 313–321.
- [153] Montes S, Alcaraz-Zubeldia M, Muriel P, et al. Striatal manganese accumulation induces changes in dopamine metabolism in the cirrhotic rat. *Brain Res* 2001; 891(1–2): 123–129.
- [154] Montes S, Alcaraz-Zubeldia M, Muriel P, et al. Role of manganese accumulation in increased brain glutamine of the cirrhotic rat. *Neurochem Res* 2003; 28(6): 911–917.
- [155] Morello M, Zatta P, Zambenedetti P, et al. Manganese intoxication decreases the expression of manganoproteins in the rat basal ganglia: an immunohistochemical study. *Brain Res Bull* 2007; 74(6): 406–415.
- [156] Morello M, Canini A, Mattioli P, et al. Sub-cellular localization of manganese in the basal ganglia of normal and manganese-treated rats. An electron spectroscopy imaging and electron energy-loss spectroscopy study. *Neurotoxicology* 2008; 29(1): 60–72.
- [157] Moreno JA, Yeomans EC, Streifel KM, et al. Age-dependent susceptibility to manganese-induced neurological dysfunction. *Toxicol Sci* 2009; 112(2): 394–404.
- [158] Moreno JA, Streifel KM, Sullivan KA, et al. Developmental exposure to manganese increases adult susceptibility to inflammatory activation of glia and neuronal protein nitration. *Toxicol Sci* 2009; 112(2): 405–415.
- [159] Moreno JA, Streifel KM, Sullivan KA, et al. Manganese-induced NF-kappaB activation and nitrosative stress is decreased by estrogen in juvenile mice. *Toxicol Sci* 2011; 122(1): 121–133.
- [160] Morganti JB, Lown BA, Stineman CH, et al. Uptake, distribution and behavioral effects of inhalation exposure to manganese (MnO₂) in the adult mouse. *Neurotoxicology* 1985; 6(1): 1–15.
- [161] Murthy RC, Srivastava RS, Gupta SK, et al. Manganese induced testicular changes in monkeys. *Exp Pathol (Jena)* 1980; 18(4): 240–244.
- [162] Nachtman JP, Tubben RE, Commissaris RL. Behavioral effects of chronic manganese administration in rats: locomotor activity studies. *Neurobehav Toxicol Teratol* 1986; 8(6): 711–715.
- [163] National Toxicology Program. NTP Toxicology and Carcinogenesis Studies of Manganese (II) Sulfate Monohydrate (CAS No. 10034-96-5) in F344/N Rats and B6C3F1 Mice (Feed Studies). National Toxicology Program Technical Report Series 428, 1–275. 1993. Technical Report Series. <http://www.ncbi.nlm.nih.gov/pubmed/12616303>.
- [164] NEHER GM, DOYLE LP, THRASHER DM, et al. Radiographic and histopathological findings in the bones of swine deficient in manganese. *Am J Vet Res* 1956; 17(62): 121–128.
- [165] Nishida M, Kawada J, Ishizuka H, et al. Goitrogenic action of manganese on female mouse thyroid through three generations. *Zoological Science* 1988; 5(5): 1043–1049.
- [166] Nishiyama K, Suzuki Y, Fujii N, et al. Biochemical changes and manganese distribution in monkeys exposed to manganese dioxide dust. *Tokushima J Exp Med* 1977; 24(3–4): 137–145.
- [167] Normandin L, Carrier G, Gardiner PF, et al. Assessment of bioaccumulation, neuropathology, and neurobehavior following subchronic (90 days) inhalation in Sprague-Dawley rats exposed to manganese phosphate. *Toxicol Appl Pharmacol* 2002; 183(2): 135–145.
- [168] Normandin L, Ann BL, Salehi F, et al. Manganese distribution in the brain and neurobehavioral changes following inhalation exposure of rats to three chemical forms of manganese. *Neurotoxicology* 2004; 25(3): 433–441.
- [169] Oner G, Senturk UK. Reversibility of manganese-induced learning defect in rats. *Food Chem Toxicol* 1995; 33(7): 559–563.
- [170] Park JD, Chung YH, Kim CY, et al. Comparison of high MRI T1 signals with manganese concentration in brains of cynomolgus monkeys after 8 months of stainless steel welding-fume exposure. *Inhal Toxicol* 2007; 19(11): 965–971.
- [171] Park RM, Bowler RM, Eggerth DE, et al. Issues in neurological risk assessment for occupational exposures: the Bay Bridge welders. *Neurotoxicology* 2006; 27(3): 373–384.
- [172] Park RM, Bowler RM, Roels HA. Exposure-response relationship and risk assessment for cognitive deficits in early welding-induced manganism. *J Occup Environ Med* 2009; 51(10): 1125–1136.
- [173] Paynter DI. Changes in activity of the manganese superoxide dismutase enzyme in tissues of the rat with changes in dietary manganese. *J Nutr* 1980; 110(3): 437–447.
- [174] Paynter DI. The role of dietary copper, manganese, selenium, and vitamin E in lipid peroxidation in tissues of the rat. *Biological Trace Element Research* 1980; 2(2): 121–135.
- [175] Peneder TM, Scholze P, Berger ML, et al. Chronic exposure to manganese decreases striatal dopamine turnover in human alpha-synuclein transgenic mice. *Neuroscience* 2011; 180: 280–292.
- [176] Pine M, Lee B, Dearth R, et al. Manganese acts centrally to stimulate luteinizing hormone secretion: a potential influence on female pubertal development. *Toxicol Sci* 2005; 85(2): 880–885.
- [177] Reeves PG, Ralston NV, Idso JP, et al. Contrasting and cooperative effects of copper and iron deficiencies in male rats fed different concentrations of manganese and different sources of sulfur amino acids in an AIN-93G-based diet. *J Nutr* 2004; 134(2): 416–425.
- [178] Reichel CM, Wacan JJ, Farley CM, et al. Postnatal manganese exposure attenuates cocaine-induced locomotor activity and reduces dopamine transporters in adult male rats. *Neurotoxicol Teratol* 2006; 28(3): 323–332.

- [179] Rivera-Mancia S, Montes S, Mendez-Armenta M, et al. Morphological changes of rat astrocytes induced by liver damage but not by manganese chloride exposure. *Metab Brain Dis* 2009; 24(2): 243–255.
- [180] Rivera-Mancia S, Rios C, Montes S. Manganese and ammonia interactions in the brain of cirrhotic rats: effects on brain ammonia metabolism. *Neurochem Res* 2012; 37(5): 1074–1084.
- [181] Roels HA, Ghyselen P, Buchet JP, et al. Assessment of the permissible exposure level to manganese in workers exposed to manganese dioxide dust. *Br J Ind Med* 1992; 49(1): 25–34.
- [182] Salehi F, Carrier G, Normandin L, et al. Assessment of bioaccumulation and neurotoxicity in rats with portacaval anastomosis and exposed to manganese phosphate: a pilot study. *Inhal Toxicol* 2001; 13(12): 1151–1163.
- [183] Salehi F, Krewski D, Mergler D, et al. Bioaccumulation and locomotor effects of manganese phosphate/sulfate mixture in Sprague-Dawley rats following subchronic (90 days) inhalation exposure. *Toxicol Appl Pharmacol* 2003; 191(3): 264–271.
- [184] Salehi F, Normandin L, Krewski D, et al. Neuropathology, tremor and electromyogram in rats exposed to manganese phosphate/sulfate mixture. *J Appl Toxicol* 2006; 26(5): 419–426.
- [185] Senturk UK, Oner G. The effect of manganese-induced hypercholesterolemia on learning in rats. *Biol Trace Elem Res* 1996; 51(3): 249–257.
- [186] Shailesh Kumar MV, Desiraju T. Effects of chronic manganese exposure on rat brain regional biogenic amines and GABA/glutamate system. *Biogenic Amines* 1992; 3(4): 227–235.
- [187] Shukakidze A, Lazriev I, Mitagvariya N. Behavioral impairments in acute and chronic manganese poisoning in white rats. *Neurosci Behav Physiol* 2003; 33(3): 263–267.
- [188] Shukla GS, Singh S, Chandra SV. The interaction between manganese and ethanol in rats. *Acta Pharmacol Toxicol (Copenh)* 1978; 43(5): 354–362.
- [189] Shukla GS, Chandra SV. Striatal dopamine turnover and L-dopa treatment after short-term exposure of rats to manganese. *Arch Toxicol* 1981; 47(3): 191–196.
- [190] Singh S, Shukla GS, Srivastava RS, et al. The interaction between ethanol and manganese in rat brain. *Arch Toxicol* 1979; 41(4): 307–316.
- [191] SMITH WE, ELLIS GH. Studies of the manganese requirement of rabbits. *J Nutr* 1947; 34(1): 33–41.
- [192] Son EW, Lee SR, Choi HS, et al. Effects of supplementation with higher levels of manganese and magnesium on immune function. *Arch Pharm Res* 2007; 30(6): 743–749.
- [193] Spadoni F, Stefani A, Morello M, et al. Selective vulnerability of pallidal neurons in the early phases of manganese intoxication. *Exp Brain Res* 2000; 135(4): 544–551.
- [194] St-Pierre A, Normandin L, Carrier G, et al. Bioaccumulation and locomotor effect of manganese dust in rats. *Inhal Toxicol* 2001; 13(7): 623–632.
- [195] Strause LG, Hegenauer J, Saltman P, et al. Effects of long-term dietary manganese and copper deficiency on rat skeleton. *J Nutr* 1986; 116(1): 135–141.
- [196] Streifel KM, Moreno JA, Hanneman WH, et al. Gene Deletion of nos2 Protects Against Manganese-Induced Neurological Dysfunction in Juvenile Mice. *Toxicol Sci* 2012; 126(1): 183–192.
- [197] Struve MF, McManus BE, Wong BA, et al. Basal ganglia neurotransmitter concentrations in rhesus monkeys following subchronic manganese sulfate inhalation. *Am J Ind Med* 2007; 50(10): 772–778.
- [198] Subhash MN, Padmashree TS. Regional distribution of dopamine beta-hydroxylase and monoamine oxidase in the brains of rats exposed to manganese. *Food Chem Toxicol* 1990; 28(8): 567–570.
- [199] Subhash MN, Padmashree TS. Effect of manganese on biogenic amine metabolism in regions of the rat brain. *Food Chem Toxicol* 1991; 29(8): 579–582.
- [200] Suzuki Y, Fujii N, Yano H, et al. Effects of the inhalation of manganese dioxide dust on monkey lungs. *Tokushima J Exp Med* 1978; 25(3–4): 119–125.
- [201] Takacs S, Szabo A, Oszlanczi G, et al. A pilot study with simultaneous recording of changes in motility and cortical electrical activity of rats during four weeks of oral manganese exposure. *Int J Environ Health Res* 2011.
- [202] Tapin D, Kennedy G, Lambert J, et al. Bioaccumulation and locomotor effects of manganese sulfate in Sprague-Dawley rats following subchronic (90 days) inhalation exposure. *Toxicol Appl Pharmacol* 2006; 211(2): 166–174.
- [203] Taylor PN, Patterson HH, Klimis-Tavantzis DJ. Manganese deficiency alters high-density lipoprotein subclass structure in the sprague-dawley rat. *The Journal of Nutritional Biochemistry* 1996; 7(7): 392–396.
- [204] Taylor PN, Patterson HH, Wolinsky I, et al. Manganese deficiency affects HDL1 and HDL2 composition in rats. *Nutrition Research* 1997; 17(7): 1155–1162.
- [205] Taylor PN, Patterson HH, Klimis-Tavantzis DJ. A fluorescence double-quenching study of native lipoproteins in an animal model of manganese deficiency. *Biol Trace Elem Res* 1997; 60(1–2): 69–80.
- [206] Thompson KH, Godin DV, Lee M. Tissue antioxidant status in streptozotocin-induced diabetes in rats. Effects of dietary manganese deficiency. *Biol Trace Elem Res* 1992; 35(3): 213–224.
- [207] Thomsen HS, Loegager V, Noergaard H, et al. Oral manganese for liver imaging at three different field strengths. *Acad Radiol* 2004; 11(6): 630–636.
- [208] Torrente M, Colomina MT, Domingo JL. Behavioral effects of adult rats concurrently exposed to high doses of oral manganese and restraint stress. *Toxicology* 2005; 211(1–2): 59–69.
- [209] Tran TT, Chowanadisai W, Lonnerdal B, et al. Effects of neonatal dietary manganese exposure on brain dopamine levels and neurocognitive functions. *Neurotoxicology* 2002; 23(4–5): 645–651.
- [210] Tran TT, Chowanadisai W, Crinella FM, et al. Effect of high dietary manganese intake of neonatal rats on tissue mineral accumulation, striatal dopamine levels, and neurodevelopmental status. *Neurotoxicology* 2002; 23(4–5): 635–643.
- [211] Ulrich CE, Rinehart W, Brandt M. Evaluation of the chronic inhalation toxicity of a manganese oxide aerosol. III. Pulmonary function, electromyograms, limb tremor, and tissue manganese data. *Am Ind Hyg Assoc J* 1979; 40(5): 349–353.
- [212] Ulrich CE, Rinehart W, Busey W, et al. Evaluation of the chronic inhalation toxicity of a manganese oxide aerosol. II. Clinical observations, hematology, clinical chemistry and histopathology. *Am Ind Hyg Assoc J* 1979; 40(4): 322–329.
- [213] Ulrich CE, Rinehart W, Busey W. Evaluation of the chronic inhalation toxicity of a manganese oxide aerosol-I. Introduction, experimental design, and aerosol generation methods. *Am Ind Hyg Assoc J* 1979; 40(3): 238–244.
- [214] Venkatakrishna-Bhatt H, Panchal GM. Gastrointestinal motility on long term oral exposure to manganese in mice – review and an experimental study. *Indian Journal of Occupational and Environmental Medicine* 2001; 6(4): 192–195.
- [215] Vezer T, Papp A, Hoyk Z, et al. Behavioral and neurotoxicological effects of subchronic manganese exposure in rats. *Environ Toxicol Pharmacol* 2005; 19(3): 797–810.
- [216] Vezer T, Kurunczi A, Naray M, et al. Behavioral effects of subchronic inorganic manganese exposure in rats. *Am J Ind Med* 2007; 50(11): 841–852.
- [217] Vitarella D, Wong BA, Moss OR, et al. Pharmacokinetics of inhaled manganese phosphate in male Sprague-Dawley rats

following subacute (14-day) exposure. *Toxicol Appl Pharmacol* 2000; 163(3): 279–285.

[218] Wassermann D, Wassermann M. The ultrastructure of the liver cell in subacute manganese administration. *Environ Res* 1977; 14(3): 379–390.

[219] Weber S, Dorman DC, Lash LH, et al. Effects of manganese (Mn) on the developing rat brain: oxidative-stress related endpoints. *Neurotoxicology* 2002; 23(2): 169–175.

[220] Welsh JJ, Narbaitz R, Begin-Heick N. Metabolic effects of dietary manganese supplementation in ob/ob mice. *J Nutr* 1985; 115(7): 919–928.

[221] Yamaguchi M, Inamoto K, Suketa Y. Effect of essential trace metals on bone metabolism in weanling rats: comparison with zinc and other metals' actions. *Res Exp Med (Berl)* 1986; 186(5): 337–342.

[222] Yang P, Klimis-Tavantzis DJ. Manganese deficiency alters arterial glycosaminoglycan structure in the Sprague-Dawley rat. *The Journal of Nutritional Biochemistry* 1998; 9(6): 324–231.

[223] Yang P, Klimis-Tavantzis DJ. Effects of dietary manganese on arterial glycosaminoglycan metabolism in Sprague-Dawley rats. *Biol Trace Elem Res* 1998; 64(1–3): 275–288.

[224] Yu IJ, Park JD, Park ES, et al. Manganese distribution in brains of Sprague-Dawley rats after 60 days of stainless steel welding-fume exposure. *Neurotoxicology* 2003; 24(6): 777–785.

[225] Zidenberg-Cherr S, Han B, Dubick MA, et al. Influence of dietary-induced copper and manganese deficiency on ozone-induced changes in lung and liver antioxidant systems. *Toxicol Lett* 1991; 57(1): 81–89.

Appendix B. Bibliographical databases searched and search terms

- IMnl electronic library reference list
- Ovid Medline/Embase and Toxline databases
- Reference lists of identified articles

Ovid MEDLINE/EMBASE search terms

- 1) *manganese¹
- 2) (manganese adj2 deficiency).ti,de,ab.

TOXLINE search terms

- 1) Exposure term “manganese” was combined using AND/OR operators with the following terms for health effects:

lung[*1]; pulmonary; fibrosis; asthma; FEV1; bronchi*; alveoli; respiratory; cough; wheeze; rhinitis; sputum; granuloma*; inflammat*; irritation; mutagen*; genotoxic*; mutation; chromosome near/1 aberration[*1]; micronuclei; cancer; neoplasm[*1]; carcinogen*; carcinoma; dermal; skin; contact near/1 dermatitis; hyperreactivity; allergy; hives; immunity; immune; GPMT; sensitization; teratogen*; reproduction; “reproductive toxicity”; toxic*; fertility; ovary; pregnancy; placenta; testes; sperm*; gonad*; prolactin; hormone[*1]; foetus; foetus; neonatal; neonate[*1]; newborn[*1]; infant[*1]; child*; offspring; neurodevelopment*; behaviour*; neurobehavior*; hyperactivity; lactation*; breastfeed*; kidney[*1]; blood; haemotoxic*; hemotoxic*; anaemia; anaemia; bone; skeletal; skeleton; osteoporosis; liver; hepatotoxic*; nephrotoxic*; cardiotox*; heart; endocrine; cytotox*; neurotoxic*; brain; spinal near/1 cord; Parkinson*; tremor*;

neuromotor; bradykinesia; cognitive; cognition; intellect*; dementia; memory; learning; neuropathy; biomonitoring; biological near/1 monitoring; absorption; distribution; metabolism; bio-transformation; excretion; accumulation; bioavailability; iron

2) Manganese near/2 deficiency.

References

- ATSDR (Agency for Toxic Substances and Disease Registry), 2000. Toxicological Profile for Manganese (Update). U.S. Department of Health and Human Services Atlanta (GA).
- ATSDR (Agency for Toxic Substances and Disease Registry), 2012. Toxicological Profile for Manganese. U.S. Department of Health and Human Services Atlanta (GA).
- Allen, B., Zeiger, E., Lawrence, G., Friedman, M., Shipp, A., 2005. Dose-response modeling of in vivo genotoxicity data for use in risk assessment: some approaches illustrated by an analysis of acrylamide. *Regul. Toxicol. Pharm.* 41(1), 6–27.
- Allison, P.D., 2004. Convergence problems in logistic regression. In: Altman, M., Gill, J., McDonald, M.P. (Eds.), *Numerical Issues in Statistical Computing for the Social Scientist*. John Wiley & Sons, Hoboken, NJ, pp. 238–252.
- Barnes, D., Dourson, M., 1988. Reference dose (RfD): description and use in health risk assessments. *Regul. Toxicol. Pharm.* 8, 471–486.
- Brown, K.G., Foureman, G.L., 2005. Concentration-time-response modelling for acute and short-term exposures. *Regul. Toxicol. Pharm.* 43 (October (1)), 45–54.
- Brown, K.G., Strickland, J.A., 2003. Utilising data from multiple studies (meta-analysis) to determine effective dose-duration levels: example: rats and mice exposed to hydrogen sulphide. *Regul. Toxicol. Pharm.* 37 (April (2)), 305–317.
- Chambers, A., Krewski, D., Birkett, N., Plunkett, L., Hertzberg, R., Danzeisen, R., Aggett, P.J., Starr, T.B., Baker, S., Dourson, M., Jones, P., Keen, C.L., Meek, B., Schoeny, R., Slob, W., 2010. An exposure-response curve for copper excess and deficiency. *J. Toxicol. Environ. Health Part B* 13 (7), 546–578.
- Chen, P., Chakraborty, S., Peres, T.V., Bowman, A.B., Aschner, M., 2014. Manganese-induced neurotoxicity: from *C. elegans* to humans (Review). *Toxicol. Res.* doi: <http://dx.doi.org/10.1039/c4tx00127c>.
- Crump, K.S., 1984. A new method for determining allowable daily intake. *Fundam. Appl. Toxicol.* 4, 85471.
- Deveau, M., Chen, C.P., Johanson, G., Krewski, D., Maier, A., Niven, K.J., Ripple, S., Schulte, P.A., Silk, J., Urbanus, J.H., Zalk, D.M., Niemeier, R.W., 2015. The global landscape of occupational exposure limits—implementation of harmonization principles to guide limit selection. *J. Occup. Environ. Hyg.* 12 (Suppl. 1), S127–S144.
- Gift, J.S., McCaughy, R., Singh, D.V., Sonawane, B., 2008. Health assessment of phosgene: approaches for derivation of reference concentration. *Regul. Toxicol. Pharm.* 51, 98–107.
- Guth, J.D., Raymond, T.S., 1996. A database designed to support dose-response analysis and risk assessment. *Toxicology* 114, 81–90.
- Haber, L., Strickland, J.A., Guth, D.J., 2001. Categorical regression analysis of toxicity data. *Comments Toxicol.* 7, 437–452.
- Health Canada, 1994. Risk Assessment for the Combustion Products of Methylcyclopentadienyl Manganese Tricarbonyl (MMT) in Gasoline. Health Canada.
- Krewski, D., Chambers, A., Stern, B., Aggett, P., Plunkett, L., Rudenko, L., 2010. Development of a copper database for exposure-response analysis. *J. Toxicol. Environ. Health A* 73 (2), 208–216.
- Milton, B., Farrell, P., Birkett, N., Krewski, D., 2016a. Modeling U-shaped exposure-response relationships for agents that demonstrate toxicity due to both excess and deficiency. *J. Risk Anal.* (Accepted 09.02.16 in press).
- Milton, B., Mattison, D.R., Krewski, D., Farrell, P.J., McGough, D., 2016b. Modeling U-shaped dose-response curves for manganese using categorical regression. *J. NeuroToxicol.* 21 Submitted March.
- Roels, H.A., Ghyselen, P., Buchet, J.P., Ceulemans, E., Lauwerys, R.R., 1992. Assessment of the permissible exposure level to manganese in workers exposed to manganese dioxide dust. *Br. J. Ind. Med.* 49 (1), 25–34.
- Sand, S., Portier, C.J., Krewski, D., 2011. A signal-to-noise crossover dose as the point of departure for health risk assessment. *Environ. Health Perspect.* 119, 1766–1774.
- Santos-Burgoa, C., Rios, C., Mercado, L.A., Arechiga-Serrano, R., Cano-Valle, F., Eden-Wynter, R.A., Texcalac-Sangrador, J.L., Villa-Barragan, J.P., Rodriguez-Agudelo, Y., Montes, S., 2001. Exposure to manganese health effects on the general population, a pilot study in Central Mexico. *Environ. Res. Sect. A* 85, 90–104.
- Schroeter, J.D., Dorman, D.C., Yoon, M., Nong, A., Taylor, M.D., Andersen, M.E., Clewell 3rd, H.J., 2012. Application of a multi-route physiologically based pharmacokinetic model for manganese to evaluate dose-dependent neurological effects in monkeys. *Toxicol. Sci.* 129 (2), 432–446.
- Strickland, J.A., Foureman, G.L., 2002. US EPA's acute reference exposure methodology for acute inhalation exposures. *Sci. Total Environ.* 288 (April (1–2)), 51–63.
- US EPA, 1988. Recommendations for and Documentation of Biological Values for Use in Risk Assessment. US EPA EPA/600/6–87/008 Available at: <http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=34855#Download>.

¹ “In databases with a controlled vocabulary this command focuses the term entered on the command line” (see <http://www.ovid.com/site/help/documentation/ospa/en/syntax.htm#operators>).

- US EPA (U.S. Environmental Protection Agency), 1993. Integrated Risk Information System (IRIS). 1. B. Reference Concentration for Chronic Inhalation Exposure (RfC). US EPA (U.S. Environmental Protection Agency) Substance Name ? Manganese (CASRN – 7439-96-5).
- US EPA (U.S. Environmental Protection Agency), 1994. Reevaluation of Inhalation Health Risk Associated with Methylcyclopentadienyl Manganese Tricarbonyl (MMT) in Gasoline. Office of Research and Development, Environmental Criteria and Assessment Office, Research Triangle Park, NC (EPA 600/R-94/062).
- (US EPA (US Environmental Protection Agency), 2006. CatReg Software User Manual (R-Version). Office of Research and Development, Washington, DC. oaspub.epa.gov=eims=eimscomm:getfile?pdownloadid=500572.
- US EPA (US Environmental Protection Agency), 2011. Exposure Factors Handbook: 2011 Edition. National Center for Environmental Assessment, Washington, DC EPA/600/R-09/052F. Available from the National Technical Information Service, Springfield, VA, and online at <http://www.epa.gov/ncea/efh>.
- WHO (World Health Organization), 2000. Manganese, Air Quality Guidelines for Europe. 2nd Edition WHO Regional Office for Europe, Copenhagen, Denmark European Series No. 91, pp. 154–156. ISBN 92 890 1358 3.