

Determination of total petroleum hydrocarbon (TPH) and polycyclic aromatic hydrocarbon (PAH) in soils: a review of spectroscopic and non-spectroscopic techniques

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3 **Determination of total petroleum hydrocarbon (TPH) and polycyclic aromatic**
4 **hydrocarbon (PAH) in soils: a review of spectroscopic and non-spectroscopic techniques**
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49 Science and Technology, Cranfield University. Dr. A. M. Mouazen holds a patent right of a
50 tractor-drawn visible and near-infrared (vis-NIR) soil sensor for online proximal soil sensing.
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Abstract

In the analysis of petroleum hydrocarbon-contaminated soils for total petroleum hydrocarbons (TPH) and polycyclic aromatic hydrocarbons (PAH), the role spectroscopic and non-spectroscopic techniques play are inseparable. Therefore, spectroscopic techniques cannot be discussed in isolation. In this report, spectroscopic techniques including Raman, fluorescence, infrared, visible-near-infrared spectroscopies, as well as mass spectroscopy (coupled to a gas chromatograph), and non-spectroscopic techniques such as gravimetric, immunoassay and gas chromatography with flame ionization detection are reviewed. To bridge the perceived gap in coverage of the quantitative applications of the vis-NIR spectroscopy in the rapid determination of TPHs and PAHs in soils, a detailed review of studies from the period 1999 – 2012 are presented. This report also highlights the strength and limitations of these techniques and evaluates their performance from the perspectives of their attributes of general applicability, namely: economic, portability, operational time, accuracy, and occupational health and safety considerations. Overall, the fluorescence spectroscopic technique had the best performance (85% total score) in comparison to others, while the gravimetric technique performed the least (60% total score). Method-specific solutions geared towards performance improvement have also been suggested.

Key words: Petroleum-Hydrocarbons; Soil; Raman spectroscopy; IR spectroscopy; Fluorescence spectroscopy; Mass spectroscopy; Vis-NIR spectroscopy

Abbreviations:

COPC, constituents of potential concern; DRS, diffuse reflectance spectroscopy; EPA, Environmental Protection Agency; FID, flame ionisation detection; GC, gas chromatography; IMA, immunoassay; IR, infrared; MLR, multiple linear regression; MSD, mass selective detector; MVA, multivariate analysis; PAH, polycyclic aromatic hydrocarbon; PLSR, partial

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3 least-squares regression; RMSECV, root-mean-square error of cross-validation; RMSEP,
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5 root-mean-square error of prediction; RMTs, rapid measurement techniques; RPD, residual
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7 prediction deviation; SEP, standard error of prediction; TPH, total petroleum hydrocarbon;
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10 Vis–NIR, visible–near-infrared
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16 1. Introduction

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18 In both the upstream and downstream sectors of the oil and gas industry, available
19 records show that spillage of crude-oil and its daughter products occurs frequently due to
20 natural and anthropogenic causes (1, 2). Crude-oil spill on land introduces petroleum-based
21 hydrocarbons (PHCs), which negatively impact on soil biological, chemical and physical
22 characteristics. Results of various environmental studies carried out in oil spill areas show
23 staggering levels of environmental pollution and adverse effects on biota due to the
24 hazardous nature of PHCs (e.g. 3–10). Prior to the remediation of the impacted media, a full
25 assessment of the impact of the PHCs on the environment and/or humans is essential to
26 identifying both the chemistry and the areal extent to which the PHCs exceed local threshold
27 limit values (TLV), and providing decision support on the appropriate remedial strategy to
28 adopt for effective clean-up of the environmental media. The hierarchical approach to risk
29 assessment (11–20) reflects the different types of data handling required at each stage in the
30 data gathering process. Whereas tier 1 risk assessment involves, but not limited to, the
31 quantitation of the total petroleum hydrocarbons (TPH) and *n*-alkanes to establish their risk-
32 based screening levels (RBSLs), tier 2 (i.e. generic quantitative risk assessment) involves the
33 analysis of the proximal composition and distribution of individual polycyclic aromatic
34 hydrocarbon (PAH) fractions and the indicator PAH compounds. Tier 3 involves a more
35 detailed investigation to determine the compound-specific biomarkers in the environmental
36 sample (21).
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3 Over the years, several spectroscopic and non-spectroscopic techniques have been
4 developed for the analysis of TPH and PAH in soil samples but, the most frequently used are
5 immunoassay (IMA), general gravimetry, laboratory-based gas chromatography (GC) with
6 flame ionisation detection (FID) or mass spectrometry (MS), infrared (IR) spectroscopy,
7 Raman spectroscopy and fluorescence spectroscopy. In recent times, though, a couple of
8 equally important innovative methods that has shown significantly reasonable potentials for
9 the measurement of TPH and PAH in oil-contaminated soils is emerging. These include field
10 portable GC/MS (22), and new generation near-infrared analysis (NIRA) with visible and
11 near-infrared (vis-NIR) spectroscopy (23 – 28). Although it was not until the recent past that
12 some IMA techniques (29, 30), field portable GC/MS systems (22), and vis-NIR
13 spectroscopy (28) were used to detect PAHs in soil samples. Before now, the laboratory-
14 based GC/MS systems, fluorescence spectroscopy and Raman spectroscopy have been used
15 for the analysis of PAH in environmental samples but, the GC/MS systems are mostly
16 preferred because of their relative selectivity and sensitivity (18, 31 – 33).

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36 It has been widely acknowledged that making informed decisions on remediation
37 requirements after an oil spill incident requires information about hydrocarbon fractions. It
38 also requires that the degree of accuracy achieved by different analytical techniques currently
39 available meet given standards. This of course has to associate with sampling resolution and
40 cost of analysis, as these are crucial factors for a successful evaluation of hydrocarbon
41 contamination in soils. This may explain the spate of recent efforts to evolve innovative
42 analytical techniques that are believed to be economical, rapid, less prone to occupational
43 hazards and capable of high sampling resolution for improved contaminant mapping and
44 refined soil remediation recommendation; even though they still complement the standard
45 analytical techniques. Unfortunately, to our knowledge, there is no review at the moment on
46 environmental diagnostic tools for TPH and PAHs in contaminated soils, which includes
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3 latest advancements on vis-NIR spectroscopic method. Thus, there is a gap in coverage of
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5 well over a decade since the vis-NIR spectroscopic method was first used to measure TPH in
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7 diesel-contaminated field-collected soil samples by Malley et al. (25). We acknowledge, no
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9 less, opinions already expressed by analysts that this might be due to a series of unanswered
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11 questions about the vis-NIR spectroscopic method concerning, for instance, the requirements
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13 in terms of accuracy indicators (e.g. residual prediction deviation (RPD) and root mean
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15 square error of prediction (RMSEP) values) from the industry and/or regulatory agencies for
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17 a method to be applied in routine applications. However, it is believed that it may be a
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19 worthwhile effort having recently published significant improvements regarding the TPH
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21 measurement accuracies of vis-NIR spectroscopic method as well as its ability to measure
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23 relative concentrations of PAH in soils all documented alongside other methods. Of course,
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25 the importance of such information to the industry and/or regulatory agencies cannot be
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27 overemphasised. It is also believed that with more research, a vis-NIR-based operating field
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29 protocol for TPH and PAHs can be developed for routine application.
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36 The objective of this report was to review the traditional spectroscopic and non-
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38 spectroscopic analytical techniques and selected rapid measurement techniques (RMTs)
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40 based on spectroscopy for measurement of TPH and the PAHs in contaminated soils.
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47 **2. Analytical techniques for petroleum hydrocarbons in soils**

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50 No doubt, analytical methods for petroleum hydrocarbons currently in use are numerous
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52 and would be all too a herculean task to exhaust in one review. This is the reason this review
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54 focused on a selected number of the most frequently used traditional methods and innovative
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56 techniques including the vis-NIR spectroscopic method to drive home the aim of this report.
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58 These methods are distinguishable by the level of analytical details they provide and their
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60 method of application, into: screening techniques, conventional non-specific methods, and

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3 methods for detailed component analysis (36), which may be field- and/or laboratory-based.
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5 As earlier stated, there are several field- and laboratory-based analytical methods for
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7 petroleum hydrocarbons currently in use but, the most frequently used methods (Table 1) are
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9 gas chromatography with flame ionization detection (GC/FID) (EPA Method 8015) or mass
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11 spectrometric detection (GC/MSD) (EPA Methods 8270 and 625), infrared (IR) spectroscopy
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13 (EPA Method 418.1), petroleum hydrocarbons by immunoassay (IMA) (EPA Methods 4030
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15 and 4035), and gravimetric TPH methods (EPA Method 1664). Others are Raman
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17 spectroscopy, fluorescence spectroscopy and vis-NIR spectroscopy.
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25 **2.1. Laboratory-based techniques**

26 *2.1.1. General gravimetry*

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29 Gravimetric methods employ an initial cold solvent extraction step and a final weight-
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31 difference step. In-between, though, there may be a further clean-up step with silica gel to
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33 remove biogenic material. If it does not involve a clean-up step, it is termed oil and grease
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35 (O&G) method but, if it does, it is termed TPH method (31). In the general gravimetric TPH
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37 method (EPA Method 1664), soil samples are uniformly graded by sieving, oven-dried at 105
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39 °C for 12 hours, and TPH compounds eluted with *n*-hexane. The liquid extract (eluate) is
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41 contacted with silica gel to remove biogenic polar materials and then evaporated. The residue
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43 is retained and weighed, and the weight difference is reported as a percent of the total soil
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45 sample on dry weight basis. Because of the presence of suspended solids, EPA Method 1664
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47 recommends using a 0.45- μ m filter (31). Being among the earliest methods developed,
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49 though obviously one of the fast declining choice methods (37), gravimetric methods have
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51 been widely used to determine TPH in contaminated soils (37). Before now, gravimetric
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53 methods were described as quick and inexpensive methods but, in a recent study (37), the
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55 long time required for complete hexane evaporation, of not less than 60 minutes, “elevates
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3 the energetic costs of the overall procedure”, and analytical losses at higher times that cause
4 negative errors are incurred. The latter limitation corroborates similar findings in earlier
5 studies (36, 38, 39). The extraction efficiency of gravimetric methods, albeit poor, is hugely
6 affected by the type of eluting solvent used (36, 40). Hexane has poor extraction efficiency
7 for higher molecular weight petroleum compounds (31), and low polarity, which causes the
8 co-extraction of natural organic matter containing multiple polar functional groups (37, 41).
9 Consequently, other chlorinated compounds like chloroform (42) as well as toluene (43) have
10 been used as liquid extractant. It is well known that both chloroform and toluene have serious
11 health implications as evident in the risk phrases published in their respective safety data
12 sheets. Additionally, gravimetric methods are non-specific since they give no information
13 about the type of hydrocarbon present (31, 37). As a result, they are not suitable for assessing
14 PAH compounds. Instead, the method is best suited for screening TPH in very oily sludges or
15 samples containing very heavy molecular-weight hydrocarbons since light hydrocarbons (<
16 C₁₅) are easily volatilized at temperatures below 70 to 85 °C during the evaporation step (31).
17 Detection limits for TPH of approximately 50 mg/kg in soils have been reported (31).
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41 2.1.2. *Infrared (IR) spectroscopy*

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43 This method harnesses the spectra of the stretching and bending vibration associated with
44 a molecule when it absorbs energy in the IR region of the electromagnetic spectrum for
45 property elucidation (31). In the electromagnetic spectrum, spectra of hydrocarbon
46 derivatives originate mainly from combinations or overtones of C-H stretching modes of
47 saturated CH₂ and terminal –CH₃ or aromatic C-H functional groups (44). In the IR region,
48 these occur within the wavenumber range of 3000 to 2900 cm⁻¹ (~3333 to 3448 nm) or at the
49 specific wavenumber of 2930 cm⁻¹ (~3413 nm) (33). Usually, as-received samples are first
50 extracted with an eluting solvent containing no C-H bonds and the eluate is contacted with
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3 silica gel to remove biogenic polar components before being subjected to IR spectrometry.
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5 The absorbance of the eluate is then measured at the specified wavenumber and compared
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7 against the calibration curve developed for the instrument. The instrument calibration
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9 standard usually is a petroleum hydrocarbon of known TPH concentration (31). The primary
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11 advantage of the IR-based methods is that they are quick, simple and inexpensive with
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13 commonly detection limits of approximately 10mg/kg in soil (31, 45).
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17 Before the advent of gas chromatographic (GC)-based methods, IR-based methods were
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19 frequently used to detect TPH in soils (e.g. 46) as it was recognised by the USEPA as an
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21 official TPH screening method such as EPA Method 418.1 (36) as well as by the ISO as in
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23 ISO/TR 11046 (47). But, following the ban on the use of Freon (also known as 1,1,2-
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25 trichlorotrifluoroethane – CFE) as an extracting solvent because of its potentials to deplete
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27 the ozone layer, the use of IR-based methods has plummeted over the years (31). Despite the
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29 ban, though, a handful of studies can be found in the open literatures on the use of IR-based
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31 methods (27, 45, 47). However, its use as a TPH measurement method is no longer supported
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33 by international standardisation; ISO for instance, has replaced ISO/TR 11046:1992 by
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35 ISO/DIS 16703:2001, which recommends the use of gas chromatography/flame ionization
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37 detection (GC/FID) after extraction with a halogen-free solvent (47). ISO/DIS 16703:2001
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39 has been updated since 2004. Apart from the limitations on its use, a major constraint of the
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41 IR-based method, according to literatures (36, 48), is the insensitivity of the technique to
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43 unsaturated components of weathered hydrocarbons not exhibiting detectable adsorption
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45 bands at the monitoring wavelength. Additionally, the use of standard hydrocarbon mixture
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47 different from the contaminating oil for prior equipment calibration invariably does not
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49 produce true contaminant concentration since different hydrocarbons respond differently to
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51 IR spectroscopy, since single hydrocarbon oil may not be suitable as a universal calibration
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53 standard (36, 48). This is because the proportion of saturated and unsaturated hydrocarbon
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3 groups varies with each oil derivative and produces correspondingly variable IR
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5 spectroscopic responses (45). The non-specificity of IR-based methods (45) also limits their
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7 suitability for PAH assessments. IR-based methods are prone to interferences; both negative
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9 and positive biases, due to the use of dissimilar calibration standards as the spilt oil, and from
10
11 spurious signals due to CH₃ groups associated with nonpetroleum sources (31). As stated
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13 previously, multivariate calibration solves the interference problem in general. The accuracy
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15 of IR-based techniques is dependent on the extraction efficiency of the extracting solvent,
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17 which in turn is affected by the type of solvent used (31, 45). Sample porosity also has a
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19 profound influence on IR signal intensity (27).
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27 *2.1.3. Gas Chromatography/Flame Ionization Detection (GC/FID)*

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29 The origin, principle and techniques of chromatography have been widely documented
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31 (e.g. 49). Succinctly, chromatography is a separation method in which a mixture is applied as
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33 a narrow initial zone to a stationary, porous sorbent and the components are caused to
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35 undergo differential migration by the flow of the mobile phase, a liquid or a gas (49). In gas
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37 chromatography, an inert carrier gas (helium, hydrogen or nitrogen) carries the gaseous
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39 mixture (or if aqueous, liquids with boiling points < 400 °C), which is to be analysed, through
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41 a capillary column onto a detector at the end of the column (31, 49), which allows better
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43 resolution of components in complex mixtures.
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48 In GC/FID method, as-received samples are first refrigerated at 4 °C until extraction, and
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50 dried either chemically (using a suitable drying agent, say anhydrous sodium sulphate) or
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52 physically, in an oven at 105 °C for 24 hours, to remove any residual moisture. TPH
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54 compounds in the dried samples are then extracted employing eluting solvents (e.g. acetone,
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56 dichloromethane, hexane or pentane), and different forms of adsorbents (e.g. silica gel,
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58 alumina or Florisil®) are used for the extract clean-up and fractionation into aliphatics and
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3 aromatics (32) prior to injection into a chromatographic column. Sample extracts are
4 introduced into the capillary column by headspace, purge-and-trap method (for volatile
5 compounds in the range C_6 to C_{25} or C_{36}) or direct injection method (for the less volatile
6 fractions). As the temperature of the column is gradually raised, TPH compounds are
7 separated according to their boiling points as they migrate towards the end of the column
8 onto the flame ionization detector. In the detector, the high-concentration effluent eluting the
9 column are trapped and ionized by burning them in a hydrogen-air or oxygen flame causing
10 the gas in the detector to conduct electric current, and the conductivity is measured by a DC-
11 powered collector electrode above the flame. The retention time of a compound prior to
12 elution from the column is typical of the species under a set of conditions and is used to
13 correlate the detector response to the amount of compound present. The detector responses in
14 a given range are then integrated to give the total concentration of hydrocarbons referred to
15 external and/or internal hydrocarbon standards (31, 49).

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34 GC/FID is mostly preferred for laboratory applications as they provide relative
35 selectivity and sensitivity (18, 31–33), and is recognised by the USEPA, BSI and ISO. The
36 EPA Method 8015 is to be used to determine TPH, BS ISO 15009:2002 is for volatile
37 aromatic and halogenated hydrocarbons, and BS ISO 16703:2004 is for the determination of
38 content of hydrocarbons in the range C_{10} to C_{40} (n-alkanes), from solids including soils and
39 wastes (50–53). GC/FID is used for both quantitative and qualitative applications including
40 the screening of environmental samples (54–56), unravelling the type and identity of fresh to
41 mildly weathered oil in environmental samples for pattern recognition of the petroleum
42 hydrocarbons (18, 19), and characterising and resolving the profile of unresolved complex
43 mixtures (UCM) in petroleum-contaminated sediments (57). The biodegradation rate constant
44 of petroleum-hydrocarbons in a contaminated site is highly variable and difficult to evaluate
45 due to variable site conditions. But, GC/FID has been used to develop a simple correlation

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3 model to estimate the bioventing degradation rate constant of gasoline in several soils without
4 having to conduct lengthy and expensive experiments (58). Detection limits for GC/FID
5 depend on the method and sample matrix with typical values of 10 mg/kg in soil (31).
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8 However, high analytical costs and operational time (21, 59), instrument calibration problems
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11 (60), effects of sample matrix (56), and impact of GC operating conditions (61) are some of
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14 the challenges of the method (see also Table 3).
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20 2.1.4. Gas Chromatography/Mass Spectrometry (GC/MS)

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22 Over the years, a couple of alternative detection techniques to the FID (Table 1) have
23 been developed for more detailed analysis of a wider range of sample matrix due to the
24 selectivity of the FID for hydrocarbons (32). The most prominently used is the mass
25 spectrometric detection (MSD) technique. The MSD basically uses the characteristic mass
26 spectra of molecular and/or fragmented ions produced after ion impact to identify compounds
27 in the sample (62). The mass spectrometer has been described as a universal detector because
28 of its versatility in the measurement of TPH, PAHs and the compound specific biomarkers
29 (CSB) for a wide variety of environmental samples (63), and is recommended by the USEPA
30 for the determination of both TPHs and PAHs (EPA Methods 8270 and 625). The popular
31 choice of mass selective detector for most environmental analysis is because of its specificity
32 and discrete monitoring capabilities; particularly when operated in the selective-ion mode
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3 major drawback of the GC/MS is that it requires volatile and thermally stable analytes; as
4
5 such, only about 10 % of organics are amenable to GC/MS analysis (68). More so, the MSD
6
7 is reported to have a lower sensitivity than the FID, because in the impact ion mode, the
8
9 respective detectors collect and measure different proportions of the generated molecular ions
10
11 (63). Quantitative chemical analysis with laboratory-based GC/MS is undoubtedly very
12
13 exhaustive but, like the GC/FID, involves lengthy and labour-intensive extraction protocols,
14
15 costly GC-based analysis, and is uneconomical in the assessment of large-scale
16
17 contamination involving dense sampling for mapping of zones requiring remediation (64).
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25 **2.2. Field-based techniques**

26 *2.2.1. Immunoassays (IMA)*

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31 IMA is a field-based immunochemical method in which antibodies are used to
32
33 selectively bind specific petroleum constituents (31). The underlying principle of the IMA
34
35 methods is variable; depending on the linked label used for response detection. The most
36
37 prominent are: enzyme-linked immunosorbent assay (ELISA), fluorescence immunoassay
38
39 and electrochemical immunoassay (ECIA).
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42
43 In the ELISA method, the response of an antibody to sorb the sample analyte in relation
44
45 to the enzyme-labelled analyte is determined by its optical density at sorption equilibrium.
46
47 The concentration of the analyte in the sample is inversely related to the optical density of the
48
49 antibody since the labelled enzyme, with high antibody affinity, is more sensitive to the
50
51 colouring agent (31). Currently, there are a number of commercially available ELISA test
52
53 kits including Ensys™ and RaPID™ assay (Strategic Diagnostics Newark, USA). Ensys™ is
54
55 less sensitive to heavier hydrocarbon components usually found in weathered oils (45).
56
57 RaPID™ assay has been used for the measurement of polycyclic aromatic hydrocarbons
58
59 (PAHs) in soil (68) and electrical transformer oil (69) but, according to a recent study (70), is
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3 prone to the problem of cross-reactivity. Cross-reactivity, which is the ability to respond to
4
5 compounds structurally similar to the analyte, affects the specificity of PAH immunoassays
6
7 and often results in biased results because PAHs are a class of structurally related compounds
8
9
10 (70). As a result, the ELISA test kits are unsuitable for risk-based studies, which involve the
11
12 assessment of PAHs in the medium. They are, however, recognised by the USEPA as official
13
14 screening methods for TPH (EPA Method 4030) and PAHs (EPA Method 4035).
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17
18 Fluorescence immunoassay is based on selective antigen-antibody binding and
19
20 fluorescence label reagents (31) and its use for the screening of aromatics in mainly water
21
22 samples is widespread (45). Regardless of the media, it is important to note that fluorescence
23
24 immunoassay also suffers the same cross-reactivity problems as the ELISA test kits as reports
25
26 show that about 15% cross-reactivities of the anti-Naphthalene antibody bound to seven
27
28 structurally related compounds are observed during the screening of Naphthalene in water
29
30 samples with fluorescence immunoassay (29). This problem is even more complex in real-
31
32 world situation since potential cross-reactants are unlimited in number and most are seldom
33
34 determinable; so all cross-reactivity values for the cross-reactants for PAH immunoassays is
35
36 all too difficult to determine (70).
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42 Redox-labelled electrochemical immunoassay (ECIA) is a direct competitive
43
44 immunoassay based on surface-immobilized anti-PAH monoclonal antibody and electro-
45
46 catalytic redox-labelled tracer recently developed for Benzo[a]pyrene measurement (30).
47
48 Although, detection limit of a 2.4 ng/mL was reported, it has been observed not sufficient for
49
50 most practical applications. Additionally, cross-reactivity is not peculiar to polyclonal
51
52 immunoassays but, is also a major challenge for monoclonal immunoassays (70) including
53
54 redox-labelled ECIA; suggesting that ECIA also suffers the same fate as both ELISA test kits
55
56 and fluorescence immunoassay. On top of these, it has been reported that immunoassay test
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3 methods are affected by soil matrix, age of weathered oil, and that their sensitivity to
4
5 hydrocarbons decreases with increasing soil clay content (31).
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10 2.2.2. *Fluorescence spectroscopy*

11
12 Fluorescence spectroscopy is a spectrochemical method of analysis in which the
13
14 molecules of the analyte emit longer wavelength radiation in less than a microsecond during
15
16 the process of relaxation to lower energy after excitation (Figure 1) by an incident shorter
17
18 wavelength radiation.
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22 Qualitative and quantitative information about the analyte is provided by the
23
24 characteristic emission spectrum produced (71). It is a field portable technique that has been
25
26 used to detect fluorescent compounds like the PAHs based on the principle that the intensity
27
28 of the emitted radiation is indicative of the relative concentration of the PAH as well as the
29
30 number of aromatic rings. Petroleum hydrocarbon molecules absorb energy in the wavelength
31
32 range of 200 to 400 nm and fluoresce in the range 280 to 500 nm with each molecule
33
34 fluorescing at a specific wavelength thereby enabling the possibility of differentiating
35
36 between the various molecular classes (72). Monocyclic aromatic hydrocarbons (MAHs)
37
38 fluoresce at lower wavelengths than the PAHs whereas the lower boiling PAHs such as
39
40 Naphthalene fluoresce at lower wavelengths than the higher boiling PAHs like
41
42 Benzo[*a*]pyrene (72). Some commonly used fluorescence spectroscopic methods include the
43
44 ultraviolet-induced fluorescence (UVIF) and Rapid Optical Screening Tool (ROST™) laser-
45
46 induced fluorescence (LIF).
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52 The UVIF employs a powerful UV lamp, which energises the hydrocarbons on
53
54 illumination thereby causing them to fluoresce. The fluorescence signal is detected with a
55
56 charge coupled device (CCD) camera, a polychromator (or a combination of both) or a
57
58 silicon intensified target (SIT) camera. Most application utilise cone penetrometer technology
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2
3 (CPT). The CPT enables continuous measurement over the subsurface media of the
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5 investigation; providing semi-quantitative measurements (22). In one report, the United
6
7 States Environmental Protection Agency (USEPA) has reported an average analysis time of
8
9 2.6 minutes and a detection limit of 3.4 mg/kg for field TPH measurement with the UVF-
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11 3100A device (73). Using the QED™ hydrocarbon analyser designed by QROS®, even lower
12
13 detection limits of 1 mg/kg in soils for petroleum fuels and oils and 0.1 mg/kg for PAHs in
14
15 soil can be achieved in a single 5-second analysis with a throughput capacity of up to 15
16
17 samples per hour (72). Currently, in the US state of North Carolina, the QED™ hydrocarbon
18
19 analyser has been approved as a replacement for the USEPA method 8015 (based on
20
21 GC/FID) for monitoring remediation of fuel spills from Leaking Underground Storage Tanks
22
23 (LUST) (72). Despite the quantitative strength of the UVIF, analysis of complex samples can
24
25 be difficult due to overlap of spectra of different luminescent compounds, and prior sample
26
27 extraction is required (22).
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34 The ROST™ LIF system is a tunable dye laser-induced fluorescence system designed as
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36 a field screening tool for detecting petroleum hydrocarbons in the subsurface (74). Unlike the
37
38 UVIF, the ROST™ LIF system uses a pulsed laser to cause fluorescence in PAH compounds
39
40 instead of UV light. The laser is transmitted through a lorry-mounted CPT probe (housing a
41
42 sapphire window) via excitation and emission optical fibres that are pushed into the ground
43
44 (74). Available TPH data showed that the ROST™ LIF system can achieve an accuracy of
45
46 89.2% with false negatives and positives put at 5.4% respectively, and a limit of detection
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48 (LOD) of 5 mg/kg in soil (74). However, using PLS regression analysis, Aldstadt et al (71)
49
50 reported sufficiently close match between predicted and measured PAHs for the technique to
51
52 be used as a screening tool for all but 3 (i.e. Acenaphthylene, Dibenzo[*a,h*]Anthracen and
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54 Naphthalene) of the 16 priority PAHs. The ROST™ LIF system is designed for qualitative
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56 applications as it can only detect the presence or absence or relative concentration of
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3 contaminants, sensitive to non-hydrocarbon compounds in the soil, and its sensitivity is
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5 affected by soil matrix (22).
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10 2.2.3. *Field portable gas chromatograph/mass spectrometry (GC/MS)*

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12 Prompted by the need to reduce costly delays associated with laboratory-based GC
13
14 systems, their portable version is emerging. Currently, a variety of portable GC/MS systems
15
16 exist including (among others) CT-1128 GC-MS (Constellation Technology Corp., USA),
17
18 HAPSITE (INFICON, USA), and EM 640 (Bruker Instruments, USA). Reported average
19
20 weight of the portable GC systems is between 16 and 60 kg and typical analysis run time is
21
22 less than 10 minutes for some models (75). The field portable GC/MS systems obviously
23
24 differ from their laboratory-based counterparts (discussed later) in terms of provision of real-
25
26 time quantification. They however, require prior sample extraction, on-site carrier gas,
27
28 considerable electrical power and ancillary equipment just like the laboratory-based GC
29
30 systems (22). Some analysts have also observed that the major problem with existing portable
31
32 GC and GC/MS instruments, especially the microchip GCs, is sensitivity (75). A trade off
33
34 appears to exist between the size and performance of the GC/MS instruments such that the
35
36 smaller the portable GC or GC/MS instrument are, the greater the sacrifice in sensitivity,
37
38 separating power, and identifying power (75). Thus, according to Harris (75), there will
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40 always be a place for the bench-top instrument for routine high-throughput, high-volume
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42 analysis.
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53 2.2.4. *Raman spectroscopy*

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55 Raman spectroscopy is a vibrational spectroscopy based on the inelastic scattering (Anti
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57 Stokes) of a monochromatic light source (Figure 2), usually from a laser source, used in
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3 assessing the vibration of Raman-active molecules such as the PAHs and for identifying
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5 species.
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8 The Raman signal is detected with a charge coupled device (CCD) camera. Applications
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10 of Raman spectroscopy have been widely reviewed (76), and Raman spectral data for several
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12 PAHs have also been widely reported (77). In general, Raman spectroscopy is one of the
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14 RMTs that have made it possible to obtain high quality spectra on a time scale that is
15
16 economical for analytical work (77). It suffices to say that Raman is both qualitative (78) and
17
18 quantitative (79) in its application. However, extreme care is needed to avoid laser alteration
19
20 of samples (80), and fluorescence contamination is often a problem with some Raman-based
21
22 systems (79). More so, in all Raman instrument, noise is present to some extent and is a
23
24 limiting factor in detection since it defines the detection limit of a particular compound (81).
25
26 It has also been observed that although the miniaturisation of the Raman instrument enhances
27
28 convenience, this has often come at a price as sensitivity; spectral range and spectral
29
30 resolution are sacrificed with possible negative consequences to materials identification and
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32 verification (81).
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41 *2.2.5. Visible and near-infrared (vis-NIR) spectroscopy*

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43 The historical perspectives, fundamental principles and practical applications of NIR
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45 spectroscopy have been widely reported in the literatures (82). In NIR spectroscopy,
46
47 absorption of energy by substances is due to overtones and combinations of fundamental
48
49 vibrations that occur in the mid infrared range (MIR) based on the stretching and bending of
50
51 bonds involving hydrogen and other atoms such as C-H, O-H, N-H and S-H chemical bonds
52
53 (82). Although, much of the early applications of NIR spectroscopy were for qualitative
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55 purposes in the foods and beverages industry (82), its earliest qualitative and quantitative
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57 applications in soil science predominantly for agricultural purposes were reported in the
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3 1980's (83). Towards the end of the same period, the spectral characteristics of hydrocarbons
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5 were reported by Cloutis (84). The spectra of hydrocarbons originate mainly from
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7 combinations or overtones of C-H stretching modes of saturated CH₂ and terminal CH₃, or
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9 aromatic C-H functional groups (44).
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12 For the purpose of this study however, only the quantitative cases regarding detection of
13
14 TPHs and PAHs in soils are reviewed. A comprehensive review of non-invasive NIR
15
16 spectroscopy involving several other processes and matrixes can be found in Workman (85)
17
18 and Schwartz et al. (86). Following Cloutis' work, by mid-1990's a fibre-optic NIR
19
20 reflectance sensor for the detection of organics, such as Benzene and Toluene, in soils has
21
22 been developed and tested (87). Consolidating on the outcome of those investigations, albeit
23
24 reportedly dismal, a small-scale study was initiated to investigate the applicability of
25
26 reflectance spectroscopy (1600 – 1900 nm) on sandy loam artificially contaminated with
27
28 motor oil (88). Two years later, a relatively more comprehensive study was conducted
29
30 involving three soil types artificially contaminated with diesel and gasoline with reported
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32 minimum detection limits of 0.1 and 0.5 % by weight respectively (89). Away from the norm,
33
34 Malley et al. (25) for the first time used NIR reflectance spectroscopy (1100 – 2498 nm) and
35
36 the stepwise multiple linear regression to predict concentrations of TPH in diesel-
37
38 contaminated soils collected from the field with low accuracy and high prediction error
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40 (Table 4). This low performance was attributed to the small number of sample set used and
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42 the inconsistency in the reference laboratory results (among others). A decade later, using
43
44 different calibration models, Chakraborty et al. (26) reported fair TPH validation R² for field-
45
46 collected intact soils from oil-spill sites with vis-NIR reflectance spectroscopy (Table 4).
47
48 They acknowledged that small number of sample set resulted in the failure to develop robust
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50 calibration models. The possibility of using vis-NIR spectroscopy (400-2500 nm) in
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52 reflectance mode as a RMT for TPH in crude oil- and diesel-spiked soil minerals (kaolinite,
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3 illite, smectite, carbonate and quartz) has also been reported by Forrester et al. (27) with
4 relatively high calibration errors for some of the minerals (Table 4). Bray et al. (28) used an
5 ordinal logistic regression technique for total PAH and benzo[*a*]pyrene predictions using vis-
6 NIR spectroscopy with good accuracy and moderate to high false positive rate at low and
7 high total PAH threshold respectively (Table 4). These results were attributed to a lack of
8 samples. In 2012, three studies were reported (90 – 92) utilising laboratory-constructed
9 hydrocarbon contaminated soil samples. Using several multivariate techniques and slightly
10 higher number of sample set, Chakraborty et al. (90) predicted with significantly improved
11 accuracy the amount of petroleum contamination in soil samples with vis-NIR spectroscopic
12 method (Table 4). Schwartz et al. (91) employed several petroleum hydrocarbons (PHCs) for
13 the simulated contamination of a total of 750 soil samples and used vis-NIR spectroscopic
14 method to predict their TPH levels using PLS with unsatisfactory accuracy but good
15 correlation results; a development they also attributed to the confirmed inter- and intra-
16 laboratory inconsistencies in reference TPH results. The third study predicted PAHs in sets of
17 artificially contaminated soils with vis-NIR diffuse reflectance spectroscopy (the second
18 study on PAH to be reported in two years) with reported reasonably high accuracy levels as
19 shown in Table 4 (92).

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44 It must be pointed out, however, that the interaction of NIR radiation with soil sample
45 produces soil spectra with fewer absorption features due to weak vibrational modes of
46 molecular functional groups, broad and overlapping bands, which make NIR spectra difficult
47 to interpret (93). This is besides the long pathlength of probe, which may decrease resolution
48 and accuracy of the vis-NIR spectroscopic method. However, intensive research is being
49 conducted today to improve the accuracy of vis-NIR spectroscopy.
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3. Integration, analysis and discussion

When comparing technologies for the analysis of TPH and/or PAH in contaminated soils, several important attributes of general applicability can be used. In this review, the attributes discussed include economic considerations, operational time, occupational health and safety, portability and accuracy.

3.1. Economic considerations

The current capital equipment cost of different TPH and/or PAH analytical devices are shown in Table 2, which shows that the laboratory-based GC systems appear to be the most expensive probably due to their size and sophistication. Although the capital equipment cost of the portable EM 640 system is relatively on the high side, the capital equipment cost of the bench-top zNose™ 4200 or portable 3000 Micro GC 1-, 2-channel systems is comparatively low (Table 2). The capital cost of the NIR-based systems is much lower than the laboratory-based GC systems and portable EM 640 model. It is about the same range as the portable IR-based system, and ExoScan 4100, but higher than those of IMA, fluorescence, portable Raman spectroscopic and GC-MS systems (Table 2). In terms of analytical cost, a standard PAH or TPH analysis by GC-based method currently costs ~£100 per sample whereas the cost by the IR-based method is ~£32 per sample in a commercial laboratory in a developing country such as Nigeria, which is equivalent to the cost for a similar analysis in most developed countries like the United Kingdom (UK) way back in 2005 (21). Currently in the UK, it is possible that this cost may have increased considering the present economic realities. Analysts have always attributed this to the constantly increasing running cost of the GC-based systems. It is known that the analytical costs for most alternative methods, such as the FS-based systems, increase as the number of samples analysed decrease as a result of the spread of the initial capital equipment cost across the number of samples in contrast to the

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3 fixed cost per sample of the reference laboratory methods (94). Consequently, the increase in
4
5 the analytical cost per sample for such methods cannot be a fair comparison to the reference
6
7 laboratory methods (94). For some other methods like the Raman and vis-NIR spectroscopic
8
9 methods, their analytical costs are not well documented at the moment due to the absence of
10
11 well-developed and standardised operating protocols, and cannot be fairly compared.
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16 17 *3.2. Operational time*

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20 Cycle time is the time it takes the analytical system to go from one analysis to the next
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22 (75), whereas analysis run time is the sum of the cycle time and the time spent preparing the
23
24 sample for analysis. Sample preparation has got both occupational health (discussed later)
25
26 and economic implications. During microcosm studies, experience shows that costly delays
27
28 can result from overhead expenses due to high analysis run time. As shown in Table 3, high
29
30 analysis run time is associated with techniques requiring lengthy initial sample preparation
31
32 particularly the laboratory-based GC techniques; suggesting that the methods are
33
34 uneconomical in the assessment of large-scale contamination involving dense sampling as
35
36 has been previously reported (64). The portable GC systems can compare favourably with the
37
38 non-invasive devices in terms of cycle time (Table 3) but, still involve time consuming
39
40 sample extraction protocols. Although current data for gravimetric method were not available
41
42 for comparison, the analysis run time for the laboratory-based IR, fluorescence spectroscopic
43
44 and sorption-based IMA methods that which also require sample preparation, is comparable
45
46 to those of the non-invasive Raman and NIR spectroscopic techniques. Table 3 shows that the
47
48 portable IR, Raman and NIR spectroscopic techniques tend to have shorter analysis run time,
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50 since little or no sample preparation is required, as compared to those methods that depend on
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52 prior sample extraction. The shorter analysis run time of the portable IR, Raman and NIR
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54 spectroscopic techniques makes them potentially better techniques for cost-effective
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3 assessment of large-scale contamination involving dense sampling, rapid decision making
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5 and accurate contaminant mapping.
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10 3.3. Occupational health and safety

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12 As stated earlier, there are threatening occupational health and safety concerns associated
13 with sample preparation. Preparing samples for TPH and/or PAH analysis with the
14 gravimetric method [42, 43], IMA ELISA test kits (68, 95), laboratory-based IR spectroscopy
15 (47), fluorescence spectroscopy (73) and GC-based methods (19) obviously involves
16 handling of hydrocarbon-contaminated soil samples and noxious chlorinated extraction
17 solvents, which exposes the user to potentially biological and chemical hazards. For instance,
18 the solvent, tetrachloroethylene, often used as a substitute for Freon 113 for extracting TPH
19 compounds for analysis with laboratory-based IR spectroscopy is a potential carcinogen
20 according to the classification of the International Agency for Research on Cancer (IARC),
21 and is also a central nervous system depressant, which finds its way into the human body
22 through inhalation and skin contact (Wikipedia, The Free Encyclopedia). Similarly,
23 dichloromethane (DCM), a liquid extractant for TPH and PAH analysis by GC-FID or GC-
24 MS, and chloroform and toluene, liquid extractants for TPH analysis by gravimetric method,
25 all have serious health implications as evident in the risk phrases contained in their respective
26 safety data sheets. Other equally dangerous chemicals (liquid or powder), which come in
27 sealed bottles or bags that the operator must be exposed to when the containers are opened up
28 during the making up of standard solution mixes for GC-based analysis are PAH and TPH
29 calibration standards, internal standards such as deuterated alkanes and PAHs, and surrogate
30 spike standards such as Squalane, *o*-Terphenyl, 2-Fluorobiphenyl, Heptamethylnonane etc.
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34 On the other hand, the portable IR devices such as the Agilent[®] 4100 ExoScan FTIR
35 (96), the vis-NIR devices, fluorescence devices such as the UVF 3100 (73) and Raman
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3 devices employ electromagnetic radiation for property elucidation. In Raman spectroscopy,
4 the electromagnetic radiation is capable of penetrating glass and plastic containers enabling
5 in-vessel analysis thereby eliminating or reducing possible exposure to hazardous chemicals
6 (97) but, operators still face possible exposure to fugitive lasers necessitating a Class 3B laser
7 warning on the devices to comply with regulations. Even though the vis-NIR and portable IR
8 devices employ particle physics in their applications, which eliminates the need for a liquid
9 extractant and concomitant exposure to chemical hazard, there is still a need, according to
10 their data sheets, for a close proximity between the sample and the detectors for better
11 instrument sensitivities. This implies that possible contact with the contaminated soil sample
12 and exposure to both biological and unknown hazards cannot be completely eliminated. In
13 the same vein, fluorescence spectroscopy employing CCD (charge-coupled device) camera
14 may still pose a radiation risk to the operator in addition to exposure to different proprietary
15 extraction and calibration solvents often used during sample preparation.
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36 3.4. Portability

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38 Considering recent innovations, it appears that there is no clear-cut distinction between
39 field- and laboratory-based techniques. Some techniques such as those based on IR, Raman
40 and fluorescence spectroscopies, and the GC-based techniques, which are laboratory-based,
41 are now also available in manageable sizes that can be deployed for field measurements and
42 are now more user friendly (Table 3). For examples, with the coming on stream of the
43 portable GC-MS devices (Table 3), and the portable IR systems such as the Agilent 4100
44 ExoScan FTIR (96) difficult circumstances can now be accessed *in situ* with relative ease. To
45 the best of our knowledge, apart from the gravimetric methods, virtually all the techniques
46 now come in field-implementable devices essentially for convenience and to also assuage the
47 cost on logistics. However, it has been reported that miniaturisation of the GC-MS (75) and
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3 Raman (81) devices results in loss of instrument performance. However, we were not able to
4 ascertain if this problem also affects the IR- and IMA-based techniques as well as the
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6 fluorescence and vis-NIR spectroscopic techniques. Field-portable devices in general, apart
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8 from being cost-effective, simple and easy to use, are meant to reduce the amount of time
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10 spent in conventional laboratory-based analysis, expedite the screening of contaminated sites
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12 to pave the way, if need be, for more detailed investigation (36).
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20 3.5. Accuracy

21
22 More often than not, the performance of the standard analytical techniques is used as a
23
24 benchmark for the innovative techniques because they are assumed to be very accurate when
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26 compared to the innovative methods. Consequently, the innovative techniques are seen to
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28 play a complementary role to the standard analytical techniques, and more so, as their data
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30 ultimately will have to be verified by the relevant standard analytical techniques such as the
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32 GC, IR spectroscopy, and general gravimetry (Table 4). The accuracy of an innovative
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34 analytical technique relative to a standard analytical method can be determined using
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36 different indicators (98). Although the root mean square error is commonly used for
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38 estimating the performance of NIR and IR techniques, it was not possible to be adopted since
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40 this was not reported while estimating the accuracy of other methods discussed in this report.
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42 The correlation coefficient, R^2 , value is seen in this report as the common denominator
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44 among other indicators so far reported by previous researchers (Table 4) and was, therefore,
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46 used to fairly compare how good or badly a particular innovative technique has been able to
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48 predict TPH and/or PAH values relative to a chosen standard analytical method. For a fair
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50 comparison, though, average values of R^2 were used, and for the vis-NIR technique, only
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52 validated values, R^2_p , were used (see Table 4). We assumed a R^2 value between 0.9 and 1.0 as
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54 an excellent correlation, R^2 value between 0.8 and 0.89 to be a good correlation, R^2 value
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3 between 0.7 and 0.79 to be a fair correlation, and R^2 value between 0.5 and 0.69 as a poor
4 correlation. Average values of R^2 deduced show that the fluorescence spectroscopic method
5 performed best with the highest average R^2 value of 0.95, which implies an excellent
6 correlation and suggests this technique to be the best to assess TPH and/or PAHs as
7 compared to the other innovative techniques. This is followed successively by the IMA-based
8 method (average $R^2 = 0.81$) and the vis-NIR method (average $R^2 = 0.78$), implying a good
9 and fair performance respectively. Raman spectroscopic method could not be fairly compared
10 because the standard analytical method used in the studies we accessed was either not stated
11 or was the high performance liquid chromatography (HPLC), which is outside the scope of
12 this review.
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29 **4. Overall performance and method-specific recommendations**

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31 Overall, Table 5 shows that most of the analytical techniques compare very well with
32 fluorescence spectroscopy topping the table, while the gravimetric method made it to the
33 bottom of the table. Fluorescence spectroscopy has excellent records in 60% of the attributes
34 but, a good operational time and a fair occupational health and safety records (Table 5),
35 which suggest that improvements should be geared towards reducing or completely
36 eliminating the need for prior sample preparation.
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45 The IMA evidently is economical, portable and has good accuracy but, has only a fair
46 record for operational time and health and safety issues (Table 5) due to dependence on
47 solvent extraction. So, like every other solvent extraction-dependent method, the IMA would
48 require (among others) an operating protocol that will be less dependent on, or independent
49 of, chlorinated extraction solvents. Although the GC-based method has 65% in the overall
50 assessment, its high accuracy is well depicted in Table 5. This low overall score obviously is
51 not unconnected with the effects of the poor health and safety records and the fair economic
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3 and operational time factors. The portable GC/MS systems undoubtedly had a positive
4 influence on the mobility of the method. However, the introduction of the portable GC/MS
5 devices did not have much positive impact on the economic (based on capital equipment
6 cost), operational time and health and safety concerns of the method. This is because the cost
7 of most portable GC/MS devices is still relatively high (Table 5), and the method is still
8 reliant on time consuming extraction protocols and the use of extraction solvents. Therefore,
9 operating protocols that encourage the use of non-chlorinated solvents without compromising
10 analyte recovery may well improve on its health and safety impacts, while instrument design
11 that eliminates completely the need for prior sample extraction will definitely enhance
12 timeliness. The gravimetric method is cheap and has an assumed excellent accuracy record,
13 because it is assumed that standard methods are reference frames against which innovative
14 methods are referred. But, this is a laboratory-based method with poor health and safety
15 records and a fair operational time (Table 5) due to the extraction step involved. Therefore,
16 the gravimetric method would require the same improvement recommended for other
17 extraction-dependent analytical methods earlier stated.

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39 The IR and vis-NIR spectroscopies performed equally well on the overall score sheet
40 (Table 5). The significantly improved records of the IR spectroscopy may be attributed to the
41 enhanced accuracy and portability of the method brought about by the introduction of the
42 portable IR devices. On the other hand, the vis-NIR spectroscopy has excellent records in
43 terms of portability and operational time, and good records in economic and health and safety
44 standpoints (Table 5). However, the vis-NIR spectroscopy has only a fair accuracy record,
45 which appears to be the least among other methods (Table 5). The reason for this is that NIR
46 prediction is based on overtones and combinations of fundamental vibrations occurring in the
47 mid IR region. Therefore, significant improvement on the accuracy of the vis-NIR
48 spectroscopy is required. Possible solutions may include: the use of non-linear multivariate
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3 analytical methods such as artificial neural networks (ANN) as an alternative to PLS
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5 regression analysis, and understanding the effects of other affecting parameters (e.g. soil
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7 texture, moisture content and oil concentration) on soil diffuse reflectance spectra of
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9 contaminated soils. Soil properties have been predicted with higher accuracy with the ANN
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11 than the PLS regression analysis with the vis-NIR spectroscopy (99). Similarly, Mouazen et
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13 al. (100) and Mouazen et al. (101) have reported that soil texture and moisture content
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15 respectively, negatively affect the performance of NIR spectroscopy in the prediction of soil
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17 chemical properties. By sufficiently understanding the individual and combined effects of
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19 these three factors, it is expected to establish techniques to reduce or even eliminate the
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21 effects of these factors and thus lead to improved accuracy of calibration models developed to
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23 predict TPH and/or PAHs in petroleum hydrocarbon-contaminated soils. Laboratory- and
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25 field-scale studies to address these key areas are being undertaken by the authors.
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33 34 **4. Conclusions**

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36 On a regular basis, analytical techniques for TPH and/or PAHs in soils are being
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38 developed and diverse factors are coalescing to bring this about. Top on this list of factors is
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40 the increasingly persistent demand for more profitable and simple environmental diagnostic
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42 tools capable of generating reliable data on a time scale. From this study, it is obvious that
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44 there is seldom an analytical technique that is problem-free although, some are less
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46 problematic than others. This is why efforts have been made in this study to extract as much
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48 relevant information from the open access literature as possible on the application of vis-NIR
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50 spectroscopy in particular, and analytical techniques in general, in the detection of petroleum
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52 contamination in soils. If timeliness and operator health and safety are anything to go by,
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54 improved protocols aimed at completely eliminating the need for prior sample preparation
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56 and the use of chlorinated extraction solvents become a consequent tandem. This suggested
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3 method change should necessarily target the gravimetric, GC-based, IMA-based, and
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5 fluorescence spectroscopic techniques because of their low performance in those two
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7 attributes due to their dependence on lengthy sample preparation steps involving the use of
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9 chlorinated extraction solvents and proprietary extraction kits. Although the vis-NIR
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11 technique compares well with other techniques in terms of cost-effectiveness, timeliness,
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13 operator health and safety and portability, it was documented to be the least accurate. This
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15 has to do with the fact that NIR prediction is based on overtones and combinations of
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17 fundamental vibrations occurring in the mid IR region. However, if the individual and
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19 interaction effects of soil texture, moisture content and oil concentration on soil diffuse
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21 reflectance spectra and calibration models developed are well understood, and if non-linear
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23 analytical techniques such as the artificial neural network (ANN) or support vector machine
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25 (SVM) and others could be applied as alternatives to linear PLS regression analysis used so
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27 far in the literature, the accuracy of the method may be improved. These are pertinent
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29 research questions that need to be answered.
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36 To this end, there always will be room for some breakthroughs in trying to improve on
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38 existing systems by scientists, and for some techniques to complement others. Therefore,
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40 there is a need for constant reviews of the progress being made to help scientists avoid
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42 duplicating efforts by re-inventing the wheel.
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9 Table 1: Summary of selected spectroscopic and non-spectroscopic techniques for petroleum
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Table 1: Summary of selected spectroscopic and non-spectroscopic techniques for petroleum hydrocarbon measurement (modified with permission from Reference (31))

Measurement technique	Detection device	Measured Target
Gas Chromatography	Flame ionisation detector (FID)	TPH
	Mass spectrometry (MS)	TPH, PAH and CSB
Infrared Spectroscopy	IR spectrometer	TPH and PAH
General Gravimetry	Gravimetric balance	TPH
Immunoassay	ELISA kits	TPH and PAH
	ECIA kits	TPH and PAH
Raman Spectroscopy	CCD detector	TPH and PAH
Fluorescence Spectroscopy	Polychromator / CCD camera	TPH and PAH
	SIT camera	TPH and PAH
Visible and Near-infrared Spectroscopy	High intensity probe / Mug lamp	TPH and PAH

CCD, Charge-coupled device

CSB, Compound specific biomarkers

ECIA, Electrochemical immunoassay

ELISA, Enzyme-linked immunosorbent assay

PAH, Polycyclic aromatic hydrocarbon

SIT – Silicon intensified target

TPH, Total petroleum hydrocarbon

Table 2: Capital cost of selected total petroleum and polycyclic aromatic hydrocarbon analytical devices (as of 2012; from the below companies †)

Technique	Make	Model	Price (\$) (approx.)
Gas Chromatography	Agilent Technologies	6890 N GC/FID	28,000 (used)
	Agilent Technologies	GC/MS 6890/5975	39,900 (used)
	Thermo Fisher	LTQ Orbitrap	362,486
	Agilent Technologies	3000 Micro GC 1-, 2-channel systems (portable)	17,000–50,000
	zNose™	Model 4200 (bench-top)	21,000–26,000
	Bruker Instruments	EM 640 (portable)	149,530–175,375
Infrared Spectroscopy	Shimadzu	IR400	3,528
	Thermo Fisher	370 DTGS	7,055
	Thermo Scientific	Nicolet 6700 FTIR	14,900 (used)
	Agilent Technologies	4100 ExoScan FTIR (Diffuse Reflectance Head)	43,497
	Agilent Technologies	4100 ExoScan FTIR (Universal System)	61,301
Immunoassay	Pierce	Easy Titer ELISA Systems	546

† LabX, Canada; Analytik Ltd., UK; O. I. Analytical Corp., USA; StellarNet Inc., USA

Table 2: Capital cost of selected total petroleum and polycyclic aromatic hydrocarbon analytical devices (as of 2012; from the below companies †)

Technique	Make	Model	Price (\$) (approx.)
Visible and Near-infrared Spectroscopy	Analytical Spectral Devices	Quality-SpecPro [®]	48,563
		LabSpec [®] 2500	56,688
		LabSpec [®] 5000	61,078
Fluorescence Spectroscopy	Hitachi	F-4010	2,500
	Tecan	Spectra Fluor	8,200
Raman Spectroscopy	Perkin Elmer	LS5B	4,500
		-	Raman-HR-TEC-IG
	-	Raman-HR-TEC	5,295
	-	Raman-HR	4,000
	-	Raman-SR	3,500

† LabX, Canada; Analytik Ltd., UK; O. I. Analytical Corp., USA; StellarNet Inc., USA

Table 3: Characteristics of selected petroleum hydrocarbon measurement techniques (modified with permission from Reference (31))

Technique	Application method	Approx. LDL (mg/kg) ^a	Estimated analysis run time (minutes)	Sampling type	Strength	Limitations
GC-Based	Laboratory and field	10	45 – 2880 ^{b,c} (Excluding extraction time, lab-based GC cycle time may be 40 minutes while some portable devices take <10 seconds)	Purge & trap, Head-space, Solvent extraction	Selectivity and high sensitivity; oil source identification; specific (MSD); quantitative and qualitative applications, portable, rapid	Laboratory-based GCs are not for compounds < C6; Non-specific (FID); specific (MSD); Expensive (capital equipment and analytical costs); Problem of co-elution; expensive; requires expertise; produces COPC

^a In soil^b Reference (45)^c Reference (21)

LDL, Laboratory detection limit

COPC, Constituents of potential concern

Table 3: Characteristics of selected petroleum hydrocarbon measurement techniques (modified with permission from Reference (31)) *Continued*

Technique	Application method	Approx. LDL (mg/kg) ^a	Estimated analysis run time (minutes)	Sampling type	Strength	Limitations
IR-Based	Laboratory and field	6.32 ^j -15.2 ^d	1 ^m (Excluding sample extraction time for lab-based bench-top device) 0.3 ^m (No sample extraction required for the portable device)	Solvent extraction (lab-based)/ Diffuse reflectance (portable type)	Quick, simple and inexpensive, portable	Non-specific; low sensitivity; analytical losses; poor extraction efficiency; quantitative application only; produces COPC
Gravimetric	Laboratory	50	-	Solvent extraction	Quick, simple and inexpensive	Non-specific; low sensitivity; analytical losses; poor extraction efficiency; quantitative application only; produces COPC

^a In soil^d Reference (73)^j Reference (94)^m Reference (96)

Table 3: Characteristics of selected petroleum hydrocarbon measurement techniques (modified with permission from Reference (31)) *Continued*

Technique	Application method	Approx. LDL (mg/kg) ^a	Estimated analysis run time (minutes)	Sampling type	Strength	Limitations
Immunoassay	Field	10-500 (under laboratory conditions, 0.1 has been achieved with trained staff) ¹	1.5 – 3 ^c	Optical density	Quick, simple, inexpensive and portable; increasingly reasonable accuracy	Non-specific; low sensitivity; only measures aromatics; quantitative application; screening only, cross-reactivity

^a In soil^c Dexsil, USA¹ Reference (68)

LDL, Laboratory detection limit

Table 3: Characteristics of selected petroleum hydrocarbon measurement techniques (modified with permission from Reference (31)) *Continued*

Technique	Application method	Approx. LDL (mg/kg) ^a	Estimated analysis run time (minutes)	Sampling type	Strength	Limitations
NIR-Based	Field	1.0 ^f	0.6 ^g – < 2 ^h (depends on the number of scans per sample)	Diffuse reflectance/transmittance spectra	Rapid, simple, inexpensive, portable, zero-solvent extraction, non-invasive, little or no sample preparation	Non-specific, matrix effect (water, soil & nature of oil), overlapping spectra, long pathlength, indirect correlation, qualitative applications, high-level chemometrics

^f Reference (25)^g Reference (23)^h Reference (25)

LDL, Laboratory detection limit

Table 3: Characteristics of selected petroleum hydrocarbon measurement techniques (modified with permission from Reference (31)) *Continued*

Technique	Application method	Approx. LDL (mg/kg) ^a	Estimated analysis run time (minutes)	Sampling type	Strength	Limitations
Fluorescence-Based	Laboratory and field	0.05 ^d	2.6 ^d (Excludes extraction time)	Emission spectra	Rapid, portable, specific, inexpensive, quantitative & qualitative applications	Qualitative applications only, sensitive to non-hydrocarbons, sensitivity is affected by soil matrix, prior sample extraction is required
Raman Spectroscopy	Laboratory and field	-	0.1 ⁱ – < 3 ^k	Emission spectra	Rapid, portable, non-invasive, inexpensive, quantitative & qualitative application, specific	Laser alteration of samples, fluorescence contamination

^a In soil^d Reference (73)ⁱ DeltaNu, Inc., USA^k Reference (101)

LDL, Laboratory detection limit

Table 4: Measurement accuracy of selected measurement techniques for petroleum hydrocarbons in contaminated soils

Innovative method	Reference method	Deployed	Measured target	Multivariate approach	No of samples	Spectral pre-processing	Wavelength range (nm)	Statistical parameters	Reference
IMA-Based	GC-MS	Field	PAH	-	52	-	-	R ² (0.61-0.68) RSD (0.3-55 %) <i>p</i> -value (<0.001)	(68)
	GC-MS	✓	PAH	-	11	-	-	R ² (0.94-0.99)	(95)
FS-Based	GC-FID	Field	PAH	-	595	-	254	R ² (0.90-0.97) (For GRO→EDRO)	(73)
	IR-S	✓	TPH	-	30	-	370-524	RSD (5-124 %)	(102)
	GC-MS	✓	PAH	-	08	-	-	R ² (0.92)	(103)
Vis-NIR spectroscopy	GC-MS	✓	PAH	-	Not stated	-	266	R ² (0.997)	(104)
	Not stated	Laboratory and Field	Oil & Grease	PLSR	17	-	1600-1900	SEP (0.13-0.26 %) (CV)	(88)
AVG – Average									

1
2
3 CV – Cross validation
4

5 EDRO – Extended diesel range organics
6

7
8 GRO – Gasoline range organics
9

10 IR-S – Infrared spectrometer
11

12 PAC – Polynuclear aromatic compound
13

14 PAH – Polycyclic aromatic hydrocarbon
15

16
17 PLSR – Partial least squares regression
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19 RSD – Relative standard deviation (SD/AVG)
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21 SD – Standard deviation
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24 SEP – Standard error of prediction (in mg/kg)
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27 Vis-NIR – Visible and Near-infrared
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Table 4: Measurement accuracy of selected measurement techniques for petroleum hydrocarbons in contaminated soils (*continued*)

Innovative method	Reference method	Deployed	Measured target	Multivariate approach	No of samples	Spectral pre-processing	Wavelength range (nm)	Statistical parameter	Reference
Vis-NIR spectroscopy	Not stated	Laboratory and Field	Oil & Grease	PLSR	>25	Apparent absorbance, Kubelka-Munk transformation, Saunderson correction, Mean value centring, MSC	800-2700	R^2_{cv} (0.968-0.998) SEP (0.116-1.04) (CV) Bias (-0.001-0.586) RMSD (0.106-0.948) (CV) (Note: Diesel data only)	(89)
	GC-FID	✓	TPH	Stepwise MLR	26	Wavelength average, First derivative, & Smoothing splines	1100-2498	R^2_p (0.68 - 0.72) SEP (0.84- 1.00) RPD (1.76- 1.82)	(25)

1
2
3 BRT – Boosted regression tree
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5
6 FTIR – Fourier transform infrared spectroscopy
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8 MLR – Multiple linear regressions
9

10 MSC – Multiplicative scatter correction
11

12 PLSR – Partial least squares regression
13

14
15 R^2 – Correlation coefficient (CV, cross validation; P, prediction)
16

17 RMSE – Root-mean-error (CV, cross validation; P, prediction, in mg/kg)
18

19 RPD – Residual prediction deviation = (SD/RMSEP)
20

21 SEP – Standard error of prediction (in mg/kg)
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23
24 TPH – Total petroleum hydrocarbon
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27 Vis-NIR – Visible and Near-infrared
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Table 4: Measurement accuracy of selected measurement techniques for petroleum hydrocarbons in contaminated soils (*continued*)

Innovative method	Reference method	Deployed	Measured target	Multivariate approach	No of samples	Spectral pre-processing	Wavelength range (nm)	Statistical parameter	Reference
Vis-NIR spectroscopy	Gravimetric	Laboratory and Field	TPH	PLSR, BRT	46	Parabolic splice, Wavelength average, First derivative, Second derivative, smoothing splines	350-2500	R^2_{cv} (0.64-0.85) R^2_p (0.42-0.64) RMSEP (0.335-0.589) RMSECV (0.303-0.436) RPD (1.35-1.94) Bias (-0.07-0.14)	(26)

AVG – Average

FNR – False negative rate

FPR – False positive rate

MLR – Multiple linear regressions

1
2
3 MSC – Multiplicative scatter correction
4

5
6 OLR – Ordinal logistic regression
7

8 PAH – Polycyclic aromatic hydrocarbon
9

10 PLSR – Partial least squares regression
11

12 R^2 – Correlation coefficient (CV, cross validation; P, prediction)
13

14
15 RMSE – Root-mean-error (CV, cross validation; P, prediction, in mg/kg)
16

17 RPD – Residual prediction deviation = (SD/RMSEP)
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19 TPH – Total petroleum hydrocarbon
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22 Vis-NIR – Visible and Near-infrared
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Table 4: Measurement accuracy of selected measurement techniques for petroleum hydrocarbons in contaminated soils (*continued*)

Innovative method	Reference method	Deployed	Measured target	Multivariate approach	No of samples	Spectral pre-processing	Wavelength range (nm)	Statistical parameter	Reference
Vis-NIR spectroscopy	FTIR	✓	TPH	PLSR	172	Not reported	400-2500	R^2_{cv} (0.81)	(27)
								RMSECV (4500-8000 mg/kg)	
	FTIR	✓	PAH	OLR	65	Not reported	350-2500	Accuracy (65.85-90.25 %)	(28)
								FPR (0.57-0.91)	
								FNR (0.03-0.13)	
	Not stated	Laboratory and Field	TPH	PLSR, MLR, Penalised Spline	68	First derivative, Discrete wavelet transform	350-2500	R^2_{cv} (0.84-0.98)	(90)
								RMSECV (3010-4791)	
								RPD (2.50-3.97)	

MLR – Multiple linear regressions

PAH – Polycyclic aromatic hydrocarbon

PLSR – Partial least squares regression

1
2
3 R^2 – Correlation coefficient (CV, cross validation; P, prediction)
4

5 RMSE – Root-mean-error (CV, cross validation; P, prediction, in mg/kg)
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8 RPD – Residual prediction deviation = (SD/RMSEP)
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10 TPH – Total petroleum hydrocarbon
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12 Vis-NIR – Visible and Near-infrared
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Table 4: Measurement accuracy of selected measurement techniques for petroleum hydrocarbons in contaminated soils (*continued*)

Innovative method	Reference method	Deployed	Measured target	Multivariate approach	No of samples	Spectral pre-processing	Wavelength range (nm)	Statistical parameter	Reference
Vis-NIR spectroscopy	FTIR	✓	TPH	PLSR,	750	SNV, MSC,	Not stated	R^2 (0.79-0.99)	(91)
				ANN		Smoothing, first derivative, second derivative, continuum removal		AVG delta (325-6187) AVG dev. (47-68 %) Max. delta (747-11494) Max. dev. (74-143 %)	
	GC-MS	✓	PAH	PLSR	150	Noise cut, Wavelength average, Maximum normalisation, First derivative, Smoothing, Baseline correction	350-2500	R^2_{cv} (0.56-0.86) R^2_p (0.89) RMSEP (0.201) RMSECV (0.144-0.484) RPD (1.52-2.79) (CV), 2.75(P)	(92)
AVG – Average									

1
2
3 MLR – Multiple linear regressions
4

5 MSC – Multiplicative scatter correction
6

7 PAH – Polycyclic aromatic hydrocarbon
8

9 PLSR – Partial least squares regression
10

11 R^2 – Correlation coefficient (CV, cross validation; P, prediction)
12

13 RMSE – Root-mean-error (CV, cross validation; P, prediction, in mg/kg)
14

15 RPD – Residual prediction deviation = (SD/RMSEP)
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17 SNV – Standard normal variate
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19 TPH – Total petroleum hydrocarbon
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21 Vis-NIR – Visible and Near-infrared
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Table 5: Attributes of general applicability for selected analytical methods for total petroleum and polycyclic aromatic hydrocarbons in soil

	Economic considerations [†]	Operational time	Occupational Health & Safety	Portability	Accuracy ^{††}	Overall score (%)
Gas Chromatography	xx	xx	x	xxxx	xxxx	65
Infrared Spectroscopy	xxx	xxx	xx	xxxx	xxxx	80
Gravimetric	xxxx	xx	x	x	xxxx	60
Immunoassay	xxxx	xx	xx	xxxx	xxx	75
Fluorescence Spectroscopy	xxxx	xxx	xx	xxxx	xxxx	85
Raman Spectroscopy	xxxx	xxxx	xxx	xxxx	-	Incomplete
Vis-NIR Spectroscopy	xxx	xxxx	xxx	xxxx	xx	80

x Poor

xx Fair

xxx Good

xxxx Excellent

[†] Based on capital equipment cost only for fair comparison

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3 †† Note: the accuracy of the standard methods is assumed excellent since they are used as benchmarks for the innovative methods but, in
4
5
6 practice this may not be so. The accuracy of the Raman spectroscopy could not be fairly compared because the standard analytical method
7
8 used in the studies we accessed was either not stated or was the high performance liquid chromatography (HPLC), which is outside the
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10 scope of this review.
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Figures

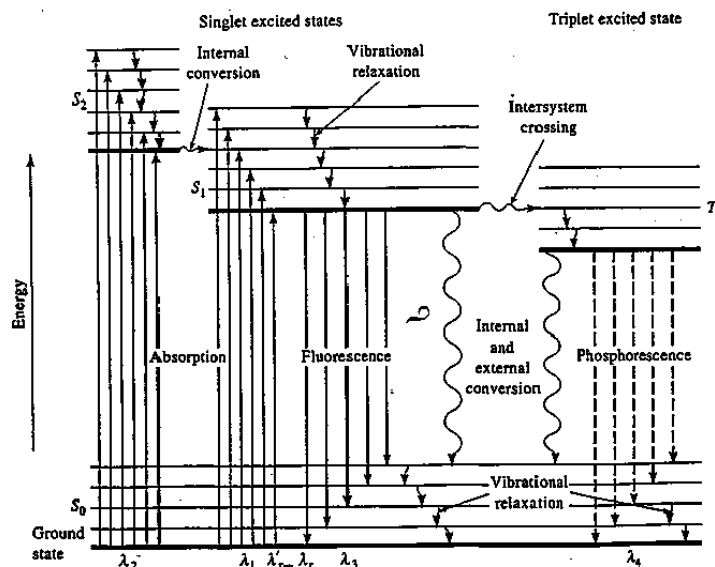


Figure 1: Electronic transition energy levels (Source: http://www.oswego.edu/~kadima/CHE425/CHE425L/FLUORESCENCE_SPECTROSCOPY_08.pdf)

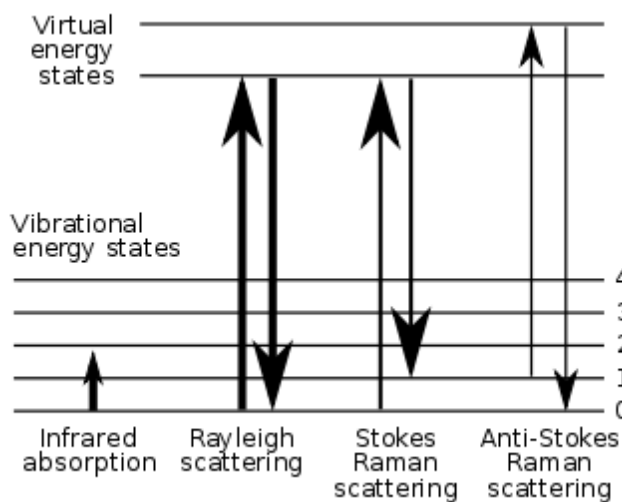


Figure 2: Energy level diagram for Raman scattering process (Source: Wikipedia, The Free Encyclopaedia: http://en.wikipedia.org/wiki/Raman_spectroscopy). The line thickness is roughly proportional to the signal strength from the different transitions.

1
2
3 **RE: Permission to use part of Table 4 (TPHCWG 1998, vol. 2)**
4

5 FROM: [Brenna Lockwood](#)

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9 Wednesday, 14 December 2011, 14:52
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11 **Message Body**

12
13 Reuben,

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15
16 You have our permission to use the table for your review paper.

17
18 Thank you,

19
20 Brenna Lockwood

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23
24 150 Fearing St., Suite 21

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26 Amherst, MA 01002

27
28 413-549-5170 T

29
30 413-549-0579 F

31
32 brenna@aehsfoundation.org

33
34 www.aehsfoundation.org

35
36
37 **From:** Reuben okparanma [mailto:rokparanma@yahoo.com]

38
39 **Sent:** Tuesday, December 13, 2011 2:21 PM

40
41 **To:** orders@aehs.com

42
43 **Subject:** Permission to use part of Table 4 (TPHCWG 1998, vol. 2)

44
45
46 Dear Editor,

47
48 I am emailing to seek permission to use part of Table 4 in pages 20 and 21 in one of your
49 publications: TPHCWG 1998, vol. 2. for a review paper intended to be submitted to the
50 Environmental Science and Technology Journal.
51

52
53 Best wishes.

54
55 Reuben.

56
57 [Reuben N. Okparanma](#),

58
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Determination of Total Petroleum Hydrocarbon (TPH) and Polycyclic Aromatic Hydrocarbon (PAH) in Soils: A Review of Spectroscopic and Nonspectroscopic Techniques

Okparanma, R. N.

2013-08-31T00:00:00Z

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